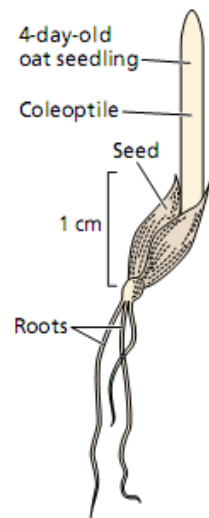
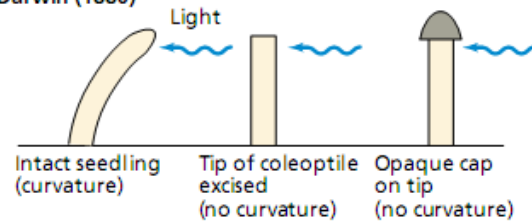


Auxins

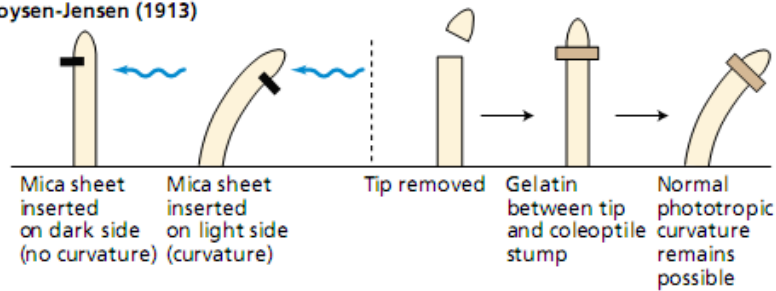


Darwin (1880)



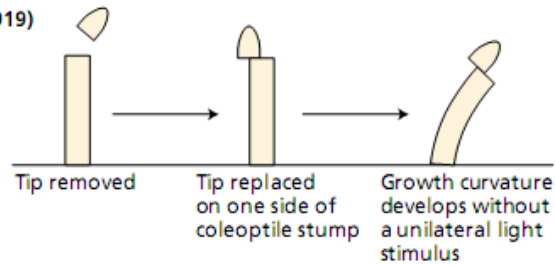
From experiments on coleoptile phototropism, Darwin concluded in 1880 that a growth stimulus is produced in the coleoptile tip and is transmitted to the growth zone.

Boysen-Jensen (1913)



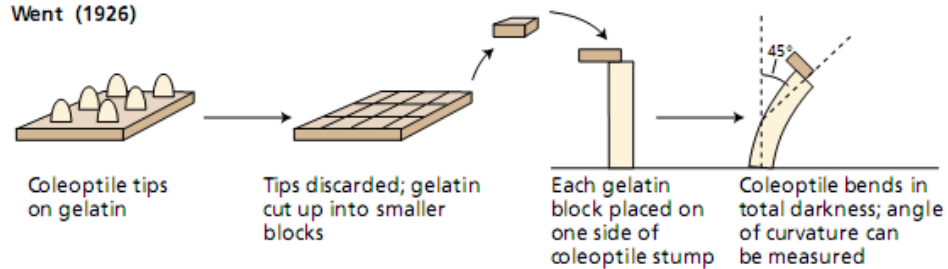
In 1913, P. Boysen-Jensen discovered that the growth stimulus passes through gelatin but not through water-impermeable barriers such as mica.

Paál (1919)

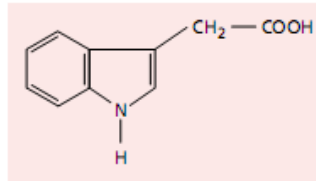


In 1919, A. Paál provided evidence that the growth-promoting stimulus produced in the tip was chemical in nature.

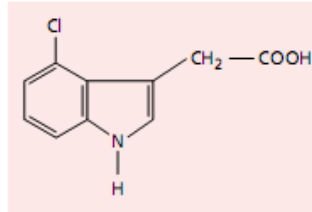
Went (1926)



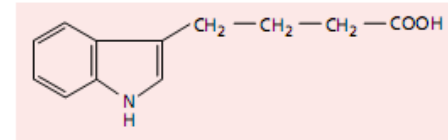
In 1926, F. W. Went showed that the active growth-promoting substance can diffuse into a gelatin block. He also devised a coleoptile-bending assay for quantitative auxin analysis.



Indole-3-acetic acid
(IAA)



4-Chloroindole-3-acetic acid
(4-Cl-IAA)



Indole-3-butyric acid
(IBA)

- 3 essential regions for auxins to bind to the auxin receptor
- 1) planar aromatic ring-acts as binding platform
- 2) a carboxylic acid acts as binding site
- 3) hydrophobic transition region-separates the 2 binding sites

Antiauxins



-
- Alpha-(p-chlorophenoxy)isobutyric acid
 - No auxin activity, competes with IAA for specific receptors without triggering auxin response

Biosynthesis

- In dividing tissues-shoot apical meristems, young leaves, developing fruits
- IPA Pathway-deamination followed by decarboxylation and then oxidation by specific dehydrogenase
- TAM pathway-deamination & decarboxylation is reversed & different enzymes are involved
- IAN pathway-cruciferae, gramineae & musaceae
- IAM pathway-indole-3-acetamide (IAM) as an intermediate--used by various pathogenic bacteria, such as *Pseudomonas savastanoi* and *Agrobacterium*

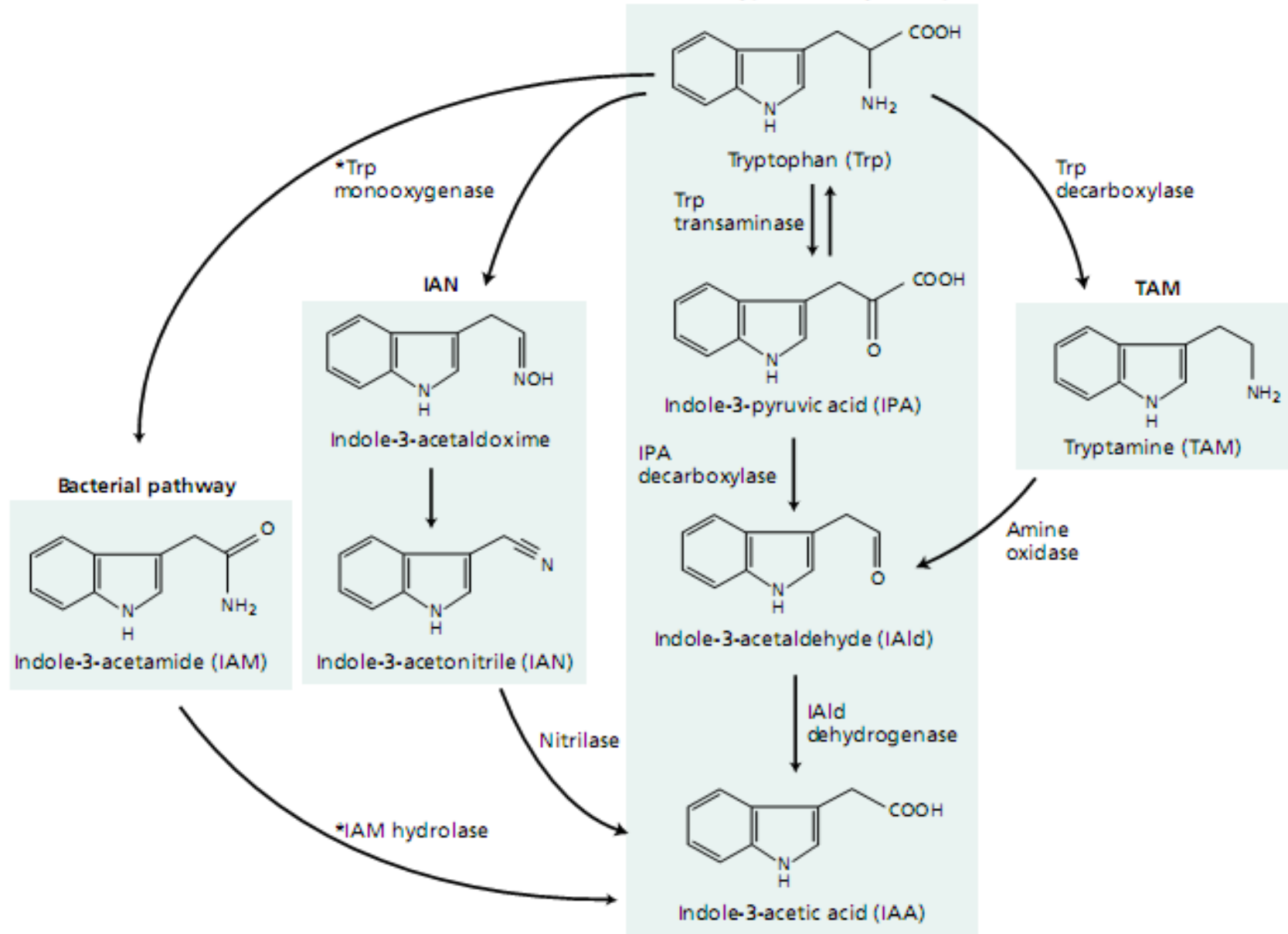
(A)

(B)

(C)

(D)

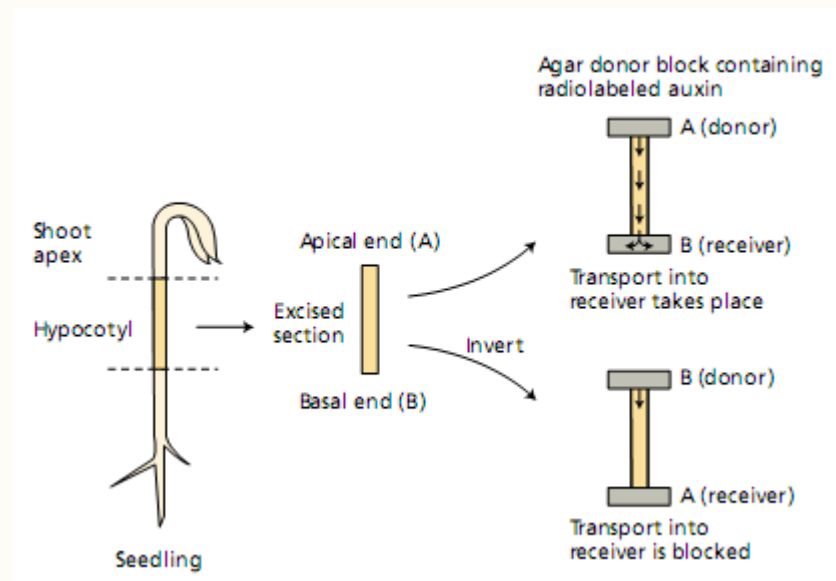
Indole-3-pyruvic acid pathway



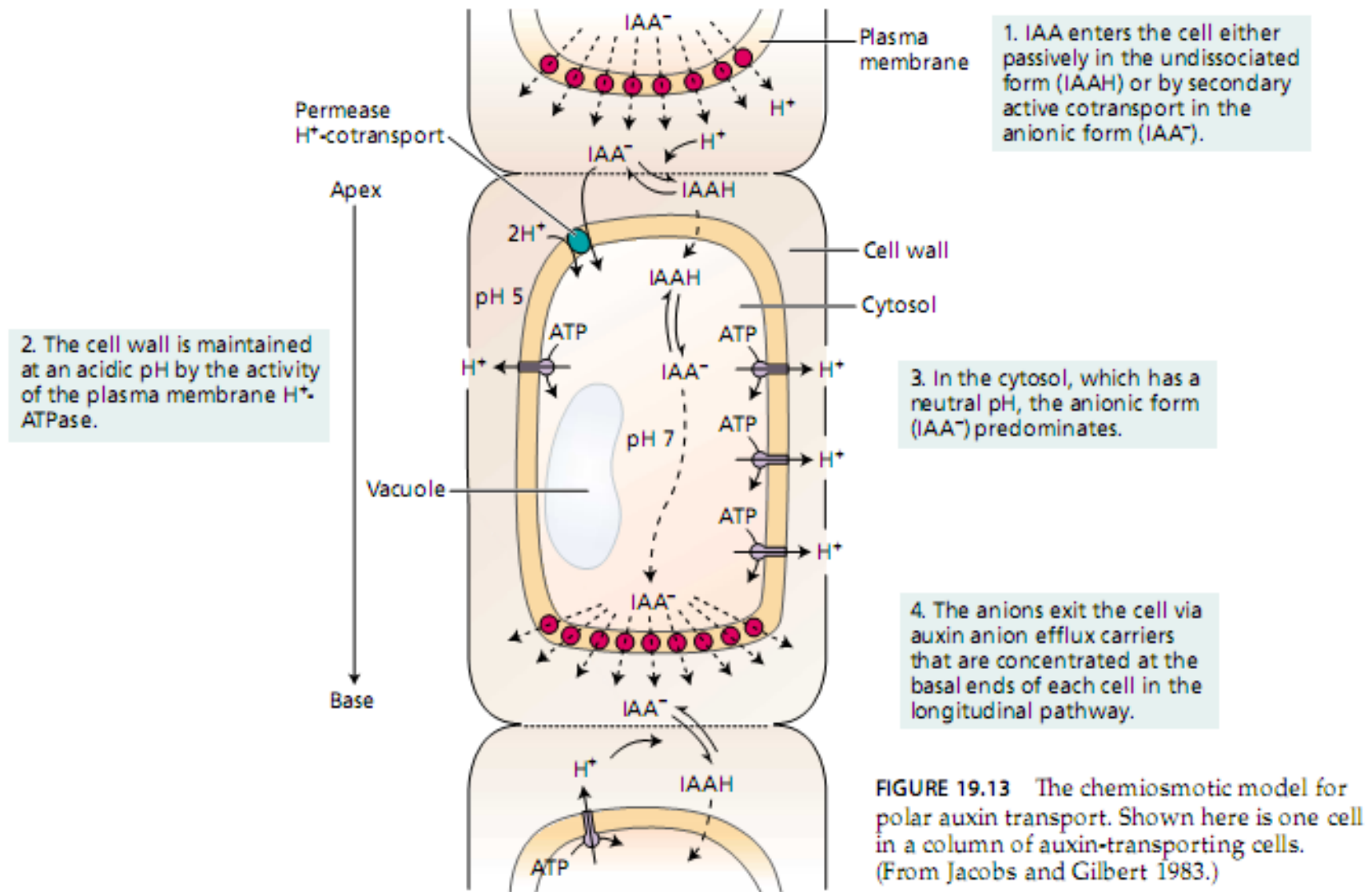



Polar transport

- Only plant growth hormone that is transported polarly. Polar transport contributes towards auxin gradient from shoot to root, which affect various developmental process, including stem elongation, apical dominance, wound healing and leaf senescence.
- Polar transport proceeds from cell to cell, rather than symplast
- High energy requiring process



A Chemiosmotic Model Has Been Proposed to Explain Polar Transport




- 
-
- Auxin efflux & influx takes place
 - The rate of polar transport is $\sim 1\text{cmh}^{-1}$ (10 times faster than diffusion)
 - Polar transport of auxins require protein carriers that can recognize the hormone (Inactive auxin analogs and auxin metabolites are not transported polarly)
 - In roots acropetal transport, small amount of basipetal auxin transport also noticed.

Inhibits Au transport

- Blocks Au efflux
- NPA(1-N-naphthylphthalamic acid)
- TIBA(2,3,5-triodobenzoic acid)
- Cause stunting, inhibition of root growth, loss of gravitropic & phototropic responses
- TIBA inhibits polar transport directly by binding to the Au binding site on the efflux carrier. In contrast phototropins bind to an associated protein that regulates the efflux carrier





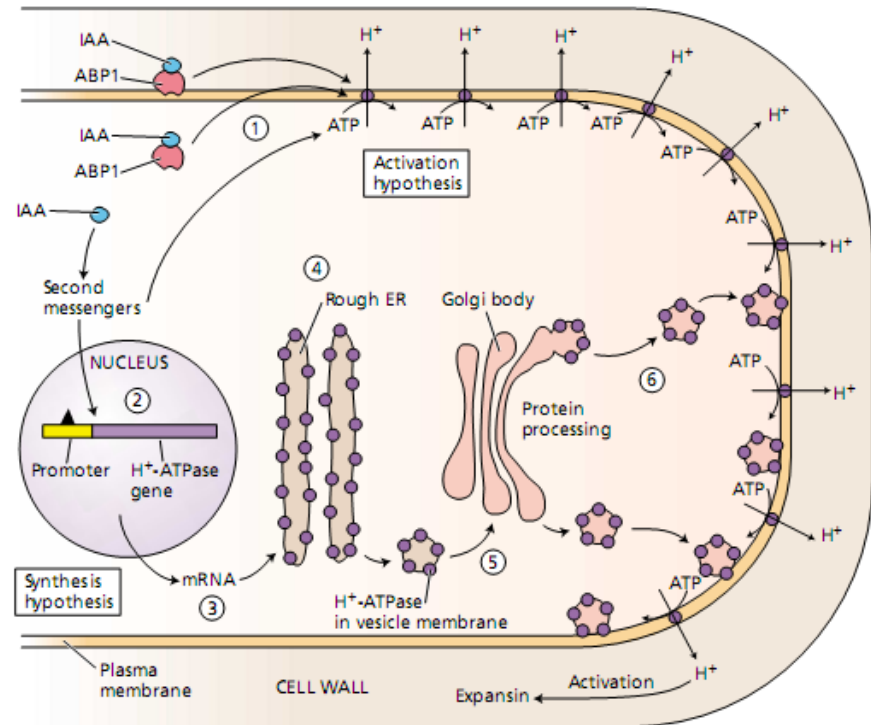
Auxin is transported nonpolarly via phloem

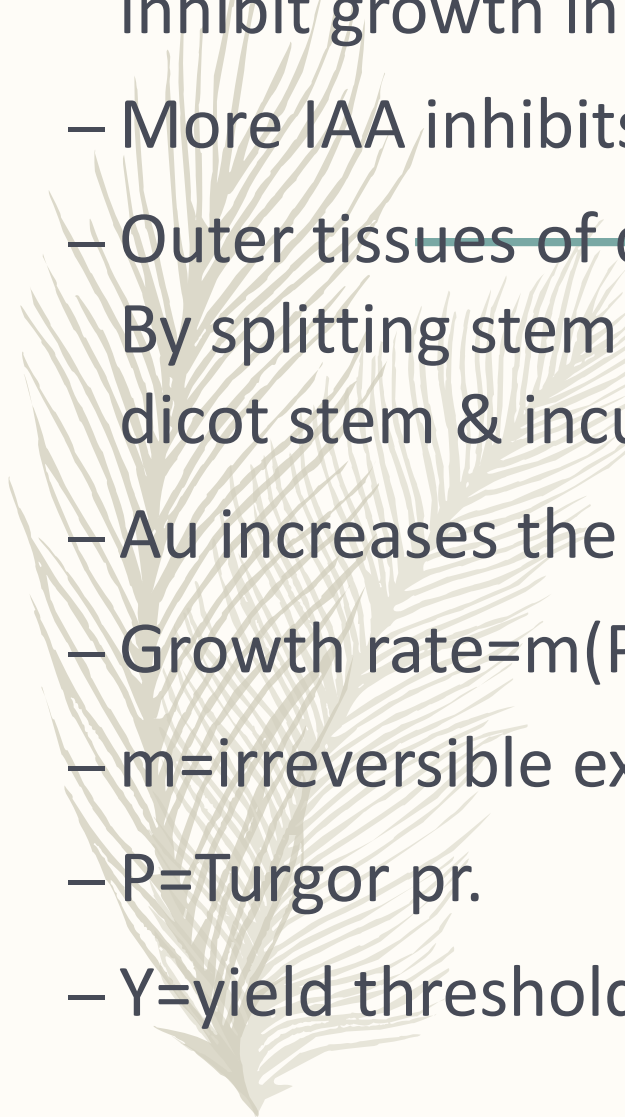
- Moves at greater speed and is passive
- Plant may be preferring phloem for long distance transport, for controlling process such as cambium division and branch root formation.
- Polar and phloem transport are not independent. Au can be transferred from the non polar phloem pathway to the polar transport pathway

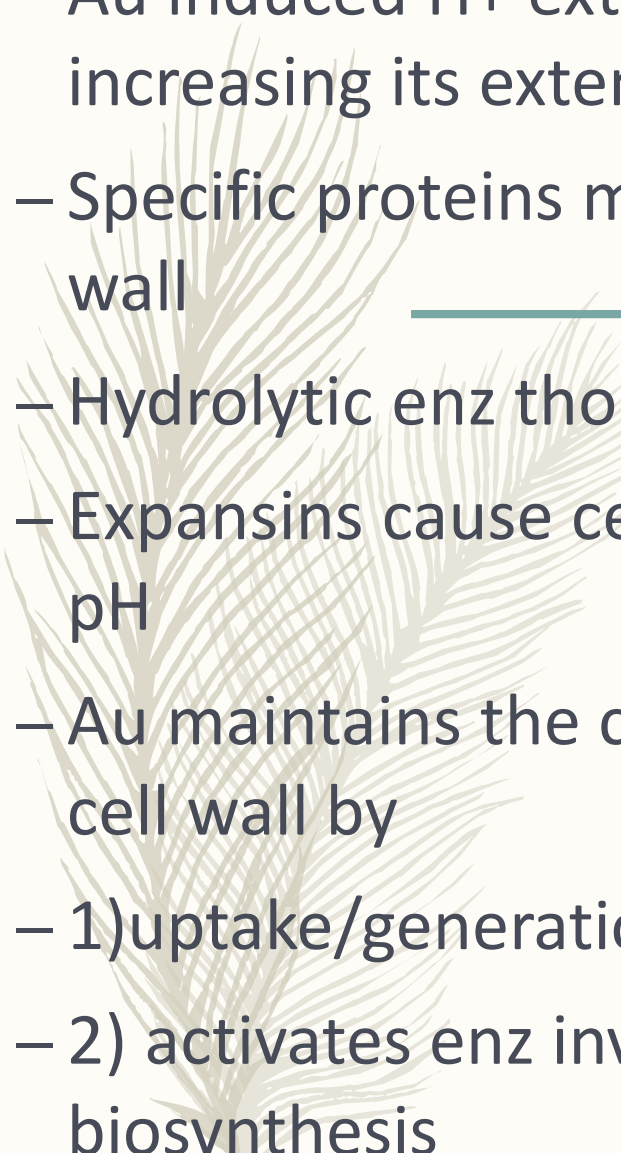
PHYSIOLOGICAL EFFECTS OF AUXIN: CELL ELONGATION

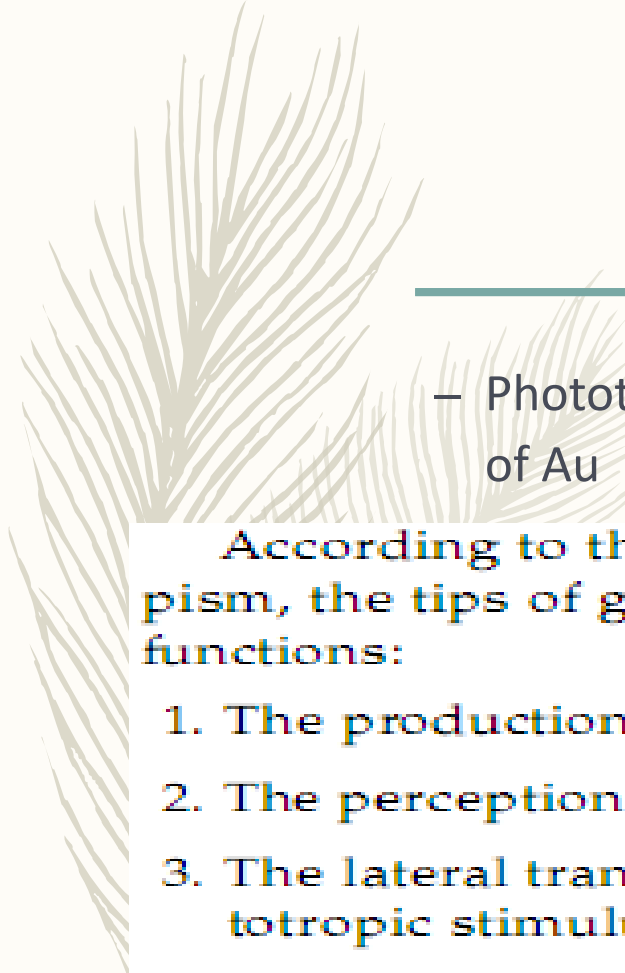
Activation hypothesis:
Auxin binds to an auxin-binding protein (ABP1) located either on the cell surface or in the cytosol. ABP1-IAA then interacts directly with plasma membrane H^+ -ATPase to stimulate proton pumping (step 1). Second messengers, such as calcium or intracellular pH, could also be involved.

Synthesis hypothesis:
IAA-induced second messengers activate the expression of genes (step 2) that encode the plasma membrane H^+ -ATPase (step 3). The protein is synthesized on the rough endoplasmic reticulum (step 4) and targeted via the secretory pathway to the plasma membrane (steps 5 and 6). The increase in proton extrusion results from an increase in the number of proton pumps on the membrane.



- 
- Au promotes growth in stems & coleoptiles but inhibit growth in roots
 - More IAA inhibits, opt is 10^{-6} to 10^{-5} M
 - Outer tissues of dicots are targets of Au action-expt.
By splitting stem sections from growing portion of dicot stem & incubating in buffer
 - Au increases the extensibility of the cell wall
 - Growth rate= $m(P-Y)$
 - m =irreversible extensibility
 - P =Turgor pr.
 - Y =yield threshold

- 
- Au induced H^+ extrusion acidifies the cell wall, increasing its extensibility: Acid growth hypothesis
 - Specific proteins mediate acid induced loosening of cell wall
-
- Hydrolytic enz thought to get activated at low pH
 - Expansins cause cell wall loosening in response to acid pH
 - Au maintains the capacity for acid induced loosening of cell wall by
 - 1) uptake/generation of osmotic solutes
 - 2) activates enz involved in cell wall polysaccharide biosynthesis




PHYSIOLOGICAL EFFECTS OF AUXIN: PHOTOTROPISM AND GRAVITROPISM

- Phototropism may be mediated by the lateral distribution of Au

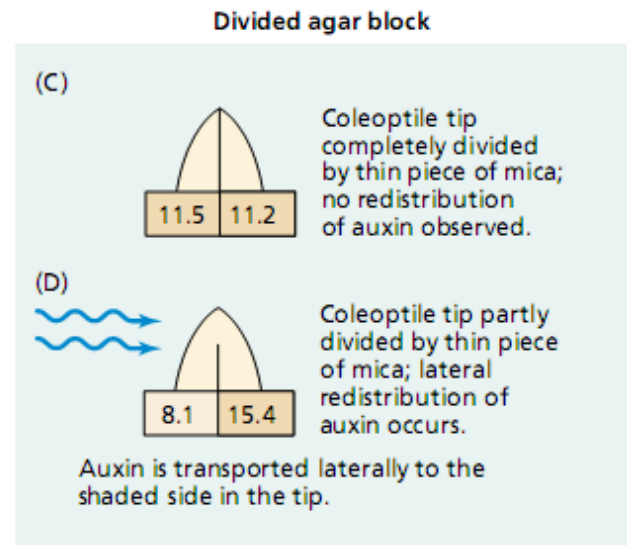
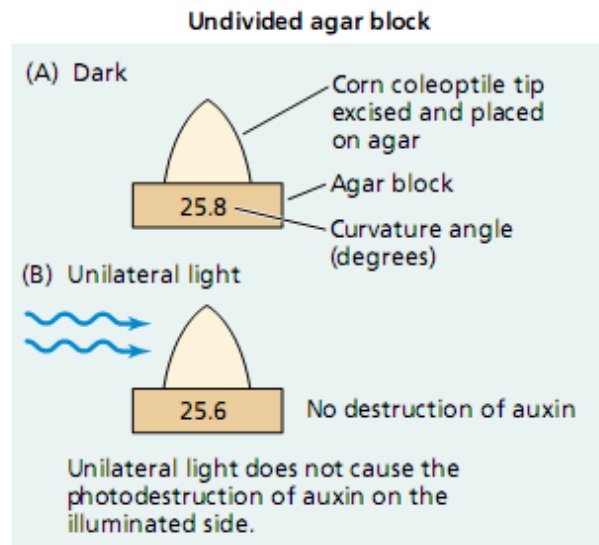
According to the Cholodny–Went model of phototropism, the tips of grass coleoptiles have three specialized functions:

1. The production of auxin
2. The perception of a unilateral light stimulus
3. The lateral transport of IAA in response to the phototropic stimulus

Thus, in response to a directional light stimulus, the auxin produced at the tip, instead of being transported basipetally, is transported laterally toward the shaded side.

- 
-
- phototropins 1 and 2, are the two flavoproteins, photoreceptors for the blue-light signaling pathway
 - Phototropin 1 displays a lateral gradient in phosphorylation during exposure to low-fluence unilateral blue light.
 - According to the current hypothesis, the gradient in phosphorylation induces the movement of Auxin to the shaded side of the coleoptile
 - Once the auxin reaches the shaded side of the tip, it is transported basipetally to the elongation zone, where it stimulates cell elongation.
 - The acceleration of growth on the shaded side and the slowing of growth on the illuminated side (differential growth) give rise to the curvature toward light.

Cholodny Went model using the agar block/coleoptile curvature bioassay



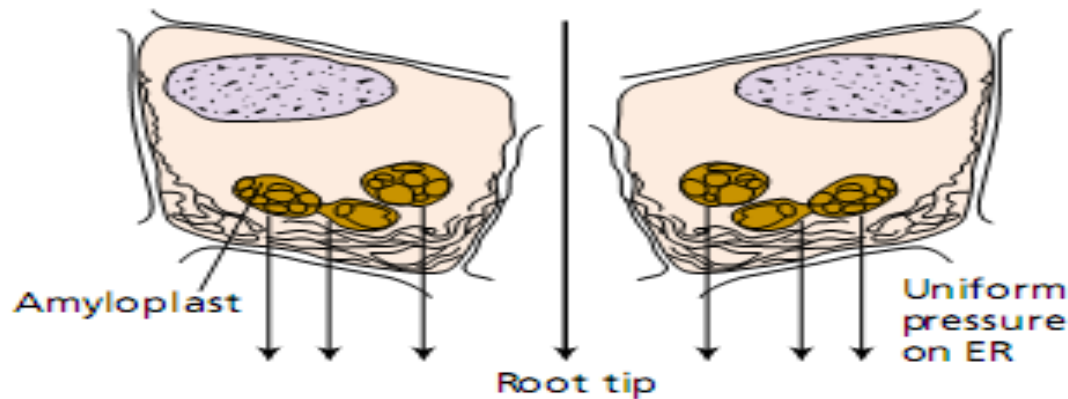


Gravitropism Also Involves Lateral Redistribution of Auxin

- Statoliths Serve as Gravity Sensors in Shoots and Roots
- Amyloplasts that function as gravity sensors are called statoliths, and the specialized gravity-sensing cells in which they occur are called statocytes.
- In shoots and coleoptiles, gravity is perceived in the starch sheath, a layer of cells that surrounds the vascular tissues of the shoot. The starch sheath is continuous with the endodermis of the root, but unlike the endodermis it contains amyloplasts.

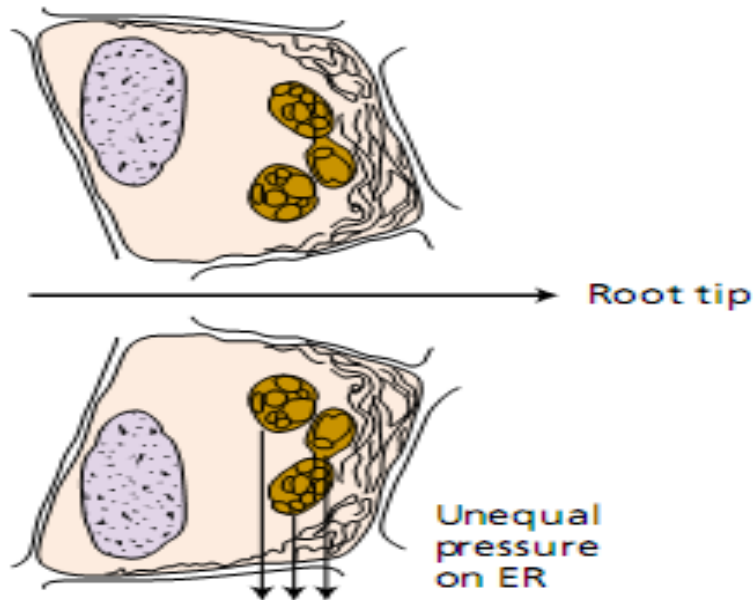
(C)

Vertical orientation



Amyloplasts tend to sediment in response to reorientation of the cell and to remain resting against the ER. When the root is oriented vertically, the pressure exerted by the amyloplasts on the ER is equally distributed.

Horizontal orientation



In a horizontal orientation the pressure on the ER is unequal on either side of the vertical axis of the root.

(A)

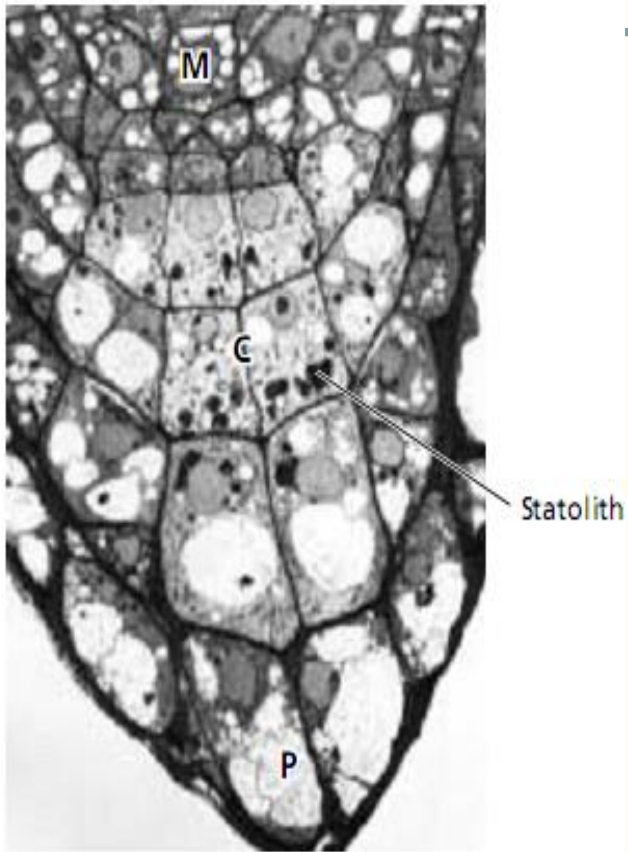
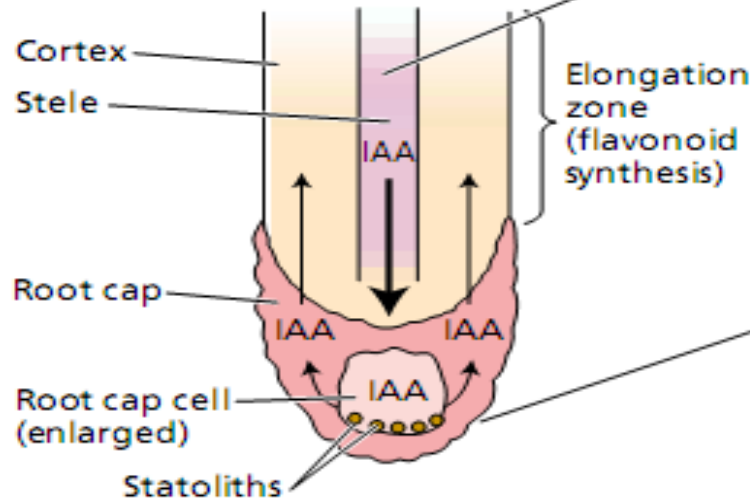


FIGURE 19.3
redistributi
pism in ma
and Evans

(A) Vertical orientation

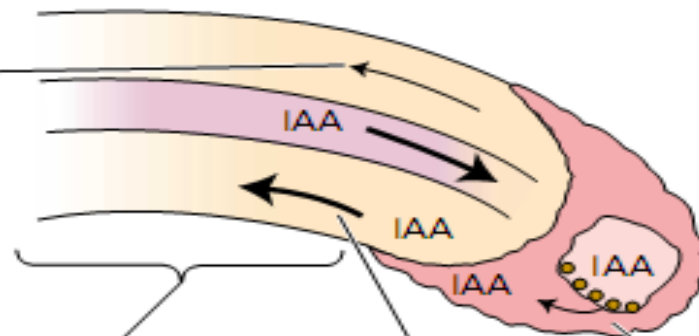


1. IAA is synthesized in the shoot and transported to the root in the stele.

2. When the root is vertical, the statoliths in the cap settle to the basal ends of the cells. Auxin transported acropetally in the root via the stele is distributed equally on all sides of the root cap. The IAA is then transported basipetally within the cortex to the elongation zone, where it regulates cell elongation.

(B) Horizontal orientation

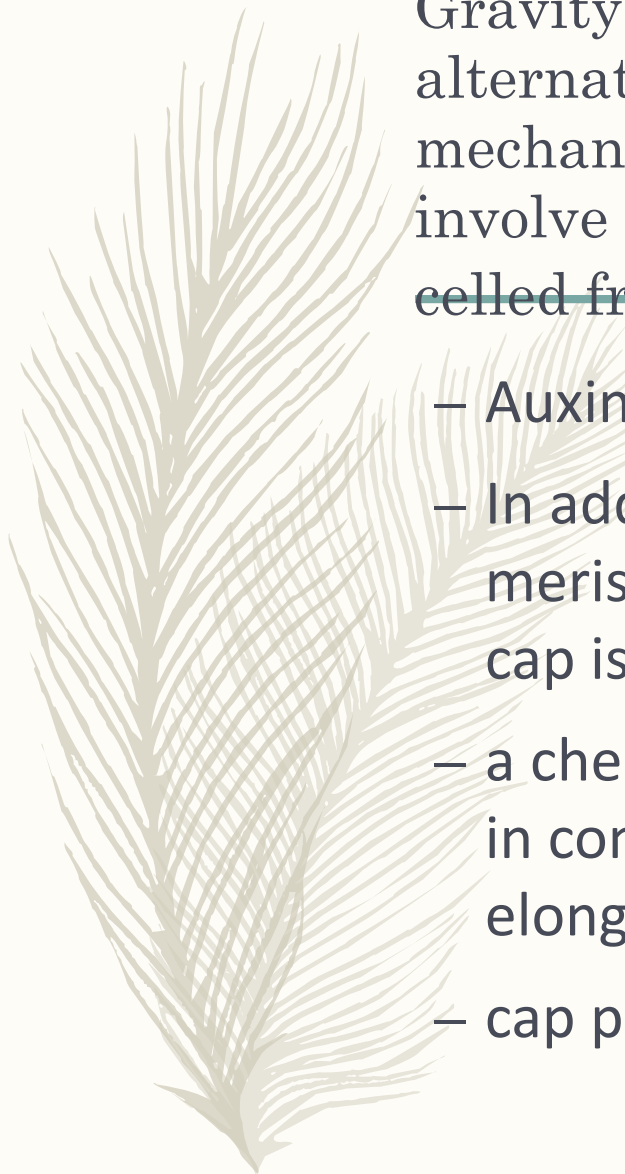
6. The decreased auxin concentration on the upper side stimulates the upper side to grow. As a result, the root bends down.



5. The high concentration of auxin on the lower side of the root inhibits growth.

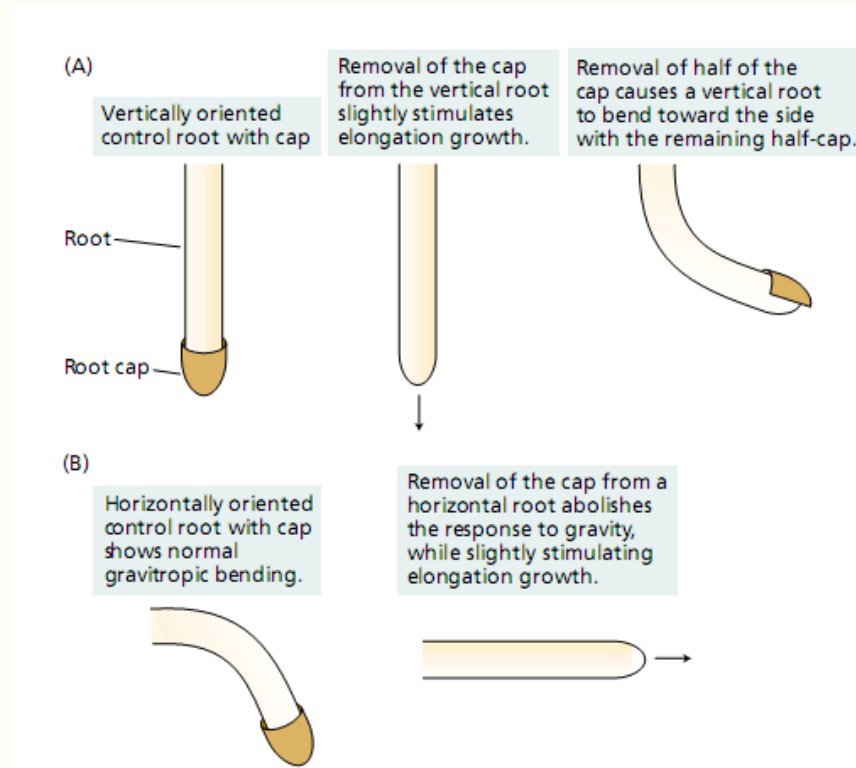
4. The majority of the auxin in the cap is then transported basipetally in the cortex on the lower side of the root.

3. In a horizontal root the statoliths settle to the side of the cap cells, triggering polar transport of IAA to the lower side of the cap.



Gravity perception without statoliths? An alternative mechanism of gravity perception that does not involve statoliths has been proposed for the giant-celled freshwater Chara.

- Auxin Is Redistribution Laterally in the Root Cap
- In addition to functioning to protect the apical meristem as the tip penetrates the soil, the root cap is the site of gravity perception.
- a chemical messenger is presumed to be involved in communication between the cap and the elongation zone.
- cap produces a root growth inhibitor (IAA)






Other physiological effects of Auxin

- Regulates apical dominance
- Promotes formation of lateral roots and adventitious roots
- Delays onset of leaf abscission
- Regulates floral bud development
- Promotes fruit development
- Induces vascular differentiation

Synthetic Au Applications

- 
- Rooting in cuttings
 - Flowering in pineapple
 - Prevention of leaf and fruit development
 - Induction of parthenocarpy
 - 2,4-D & dicamba-commonly used



Molecular mechanism of Auxin action

- Au binds to the receptors at 3 locations 1)ER (ABP1-22kDa, dimer) 2) plasma membrane 3) tonoplast
- Au binding to ABP1 increases membrane voltage
 - Ab, blocking H⁺ATPase block hyperpolarization response of Au
 - some ABP1 Ab mimics the effect of Au by bringing about hyper polarization of tobacco mesophyll protoplasts, stimulation of H⁺ extrusion etc.



Signaling intermediates in Au action

- In cell cycle, Au stimulates synthesis of cyclin dependent protein kinase (CDK)
- CDK's together with their regulatory subunits, the cyclins regulate the transitions from G1 to S and from G2 to mitosis during cell cycle.
- MAPkinases that play a role in signal transduction by phosphorylating proteins in a cascade that ultimately activates transcription factors have also been implicated in auxin responses. (when tobacco cells are deprived of auxin, they arrest at the end of the G1 or the G2 phase and cease dividing; if auxin is added back into the culture medium, the cell cycle resumes)

Contd.....



-
- Ca^{2+} (2,4-D, free Ca^{2+} rose from 280-380nm)
 - Auxin decreases cytosolic pH by 0.2 units, which in turn can promote the activity of H^{+} -ATPase, which is active under low pH
 - Auxin stimulates phospholipase A2 (PLA2)
 - fatty acids released from membrane phospholipids by PLA2 serve as substrate for lipoxygenase—the other product of the reaction lysophosphatidylcholine, can activate protein kinase (PLA2 pathway in Auxin action)

Auxin alters gene expression



-
- Rapid changes in the levels of translatable mRNA
 - Au induced genes fall into 2 classes
 - primary responsive genes (on binding of Au to receptor there is the activation of select group of preexisting transcription factors)
 - pri responsive genes have 3 main functions
 - 1) encodes protein that regulate transcription of sec response genes
 - 2) involved in cell-cell signalling
 - 3) involved in stress adaptation
 - 5 major class of early Au responsive genes
 - Aux/IAA gene family, SAUR(small Au upregulated RNA's), GH3, ACC synthase, Glutathione-S-transferase (GST's)(a class of proteins stimulated by stress conditions)

Cytokinins

Cytokinins

Discovery

1913-G.Haberlandt-austria-Vas. Tissue contains water soluble substance, stimulating the cell division in potato

Many substances tested to initiate and sustain cell poliferation – yeast extract, tomato juice, coconut milk (zeatin)

Kinetin – 1st cytokinin discovered, discovered as a DNA breakdown product

1940-50 – Skoog (Univ. Wisconsin)

Adenine (Aminopurine) – promotive effect

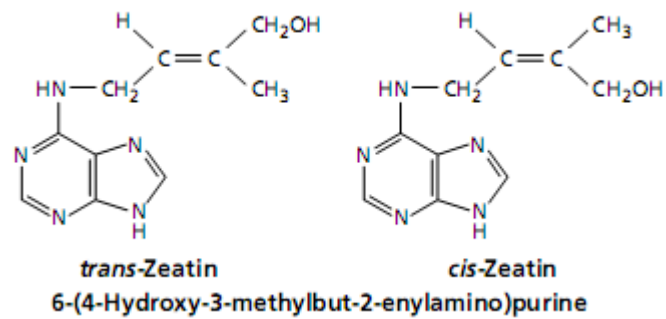
Autoclaved sperm DNA – promoting effect

Skoog & Miller – fractionating heat treated DNA – kinetin – adenine derivative (6-furfurylaminopurine)

Zeatin - abundant natural cytokinin, from extracts of immature endosperm of zeamays- (Miller & Letham)

Cis & trans forms (regulated by isomerases)

All cytokinin compounds – N6 substituted aminopurines- ex. BAP, diphenyl urea





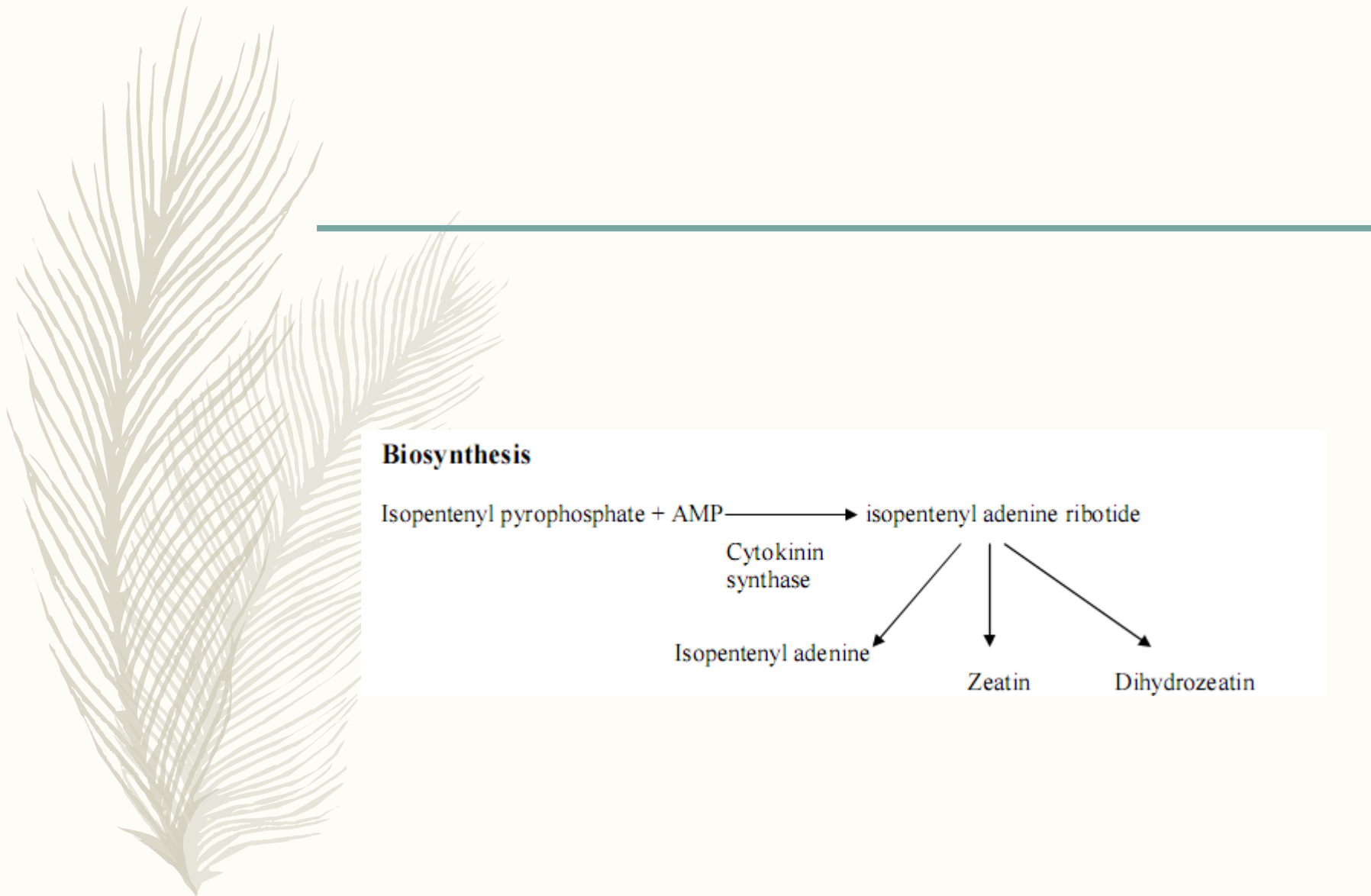
Bioassay

- 1) cell proliferation bioassay using tobacco pith tissue
- 2) cotyledonary expansion - - radish

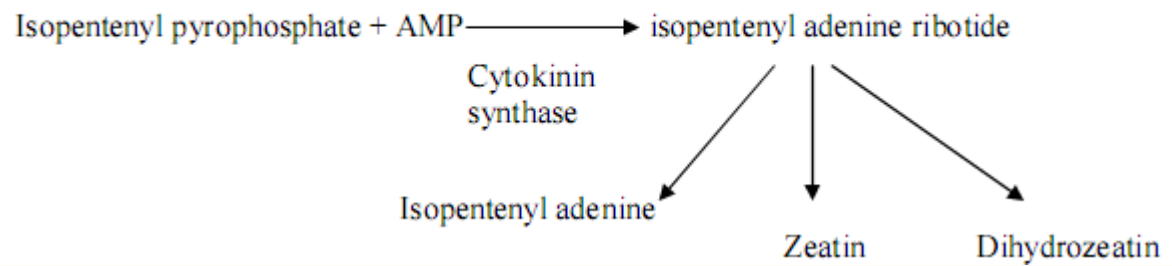
Cytokinin occur in both free and bound forms

Free- zeatin, dihydrozeatin & isopentenyladenine

Bound - plant tRNA's contain zeatin as hyper modified base



Biosynthesis



Transport

Root to shoot via xylem along with water and minerals

major site of synthesis - Root & shoot meristem, Young maize embryos, young developing leaves.

Transported as zeatin ribosides, once they reach leaf they are freed

A signal from the shoot can regulate the cytokinin transport from root.

Cytokinins are rapidly metabolized by plant tissues Glucosidases

Cytokinin bases \longrightarrow nucleotides/glucosides \longrightarrow Active form
(Storage forms, metabolically inactive) (germination initiated)
-in dormant seed

Zeatin, Zeatin riboside \longrightarrow adenine or its derivatives
Cytokinin oxidase (inactivates hormones, thus regulates it)

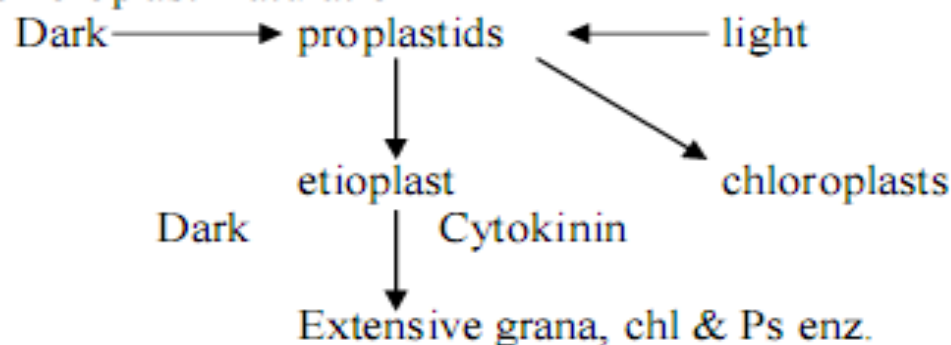


Biological roles

- 1) shoot apical meristems of cytokinin overproducing plants produce more leaves
- 3) >Chl
- 4) Adv. Shoots from leaf veins and petioles
- 5) Leaf senescence retarded
- 6) Apical dominance reduced
- 7) Cytokinin overproducing plants - stunted & shortened internodes
- 8) Rooting of stem cuttings reduced
- 9) Aux/cytokinin ratio regulates morphogenesis in cultured tissues
- 10) Modify apical dominance and promote lateral bud growth
- 11) Aux/cytokinin regulates plant cell cycle

CDK's (cyclin-the regulatory subunit is regulated by cytokinin)

- 12) delays leaf senescence (controlled by root derived cytokinins to leaves)
- 13) promotes nutrient mobilization (metabolism of treated area stimulated, so nutrients move to that side – creates a new source-sink relationship)
- 14) promotes chloroplast maturation



- 15) promotes cell expansion in leaves and cotyledons (increase in mechanical extensibility of cell walls, not accompanied by H^+ extrusion)
- 16) inhibits cell elongation in stems and roots

Tobacco plants overexpressing the gene for cytokinin oxidase. Shoot growth is strongly inhibited in the transgenics



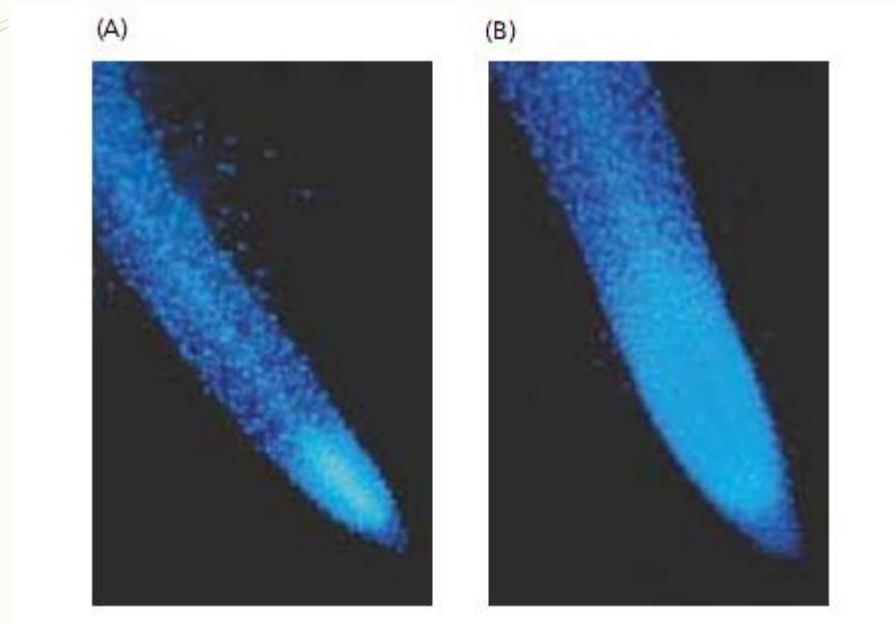
Transgenics overproducing cytokinin oxidase



Cytokinin suppresses the growth of roots.

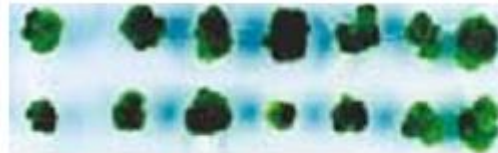


Cytokinin suppresses the size and cell division activity of roots. (A) Wild type. (B) AtCKX1.



The Auxin: Cytokinin Ratio Regulates Morphogenesis in Cultured Tissues

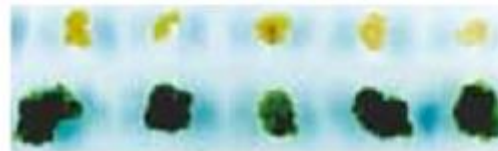
Auxin + cytokinin



wild type

CYCD3
overexpressor

Auxin



wild type

CYCD3
overexpressor

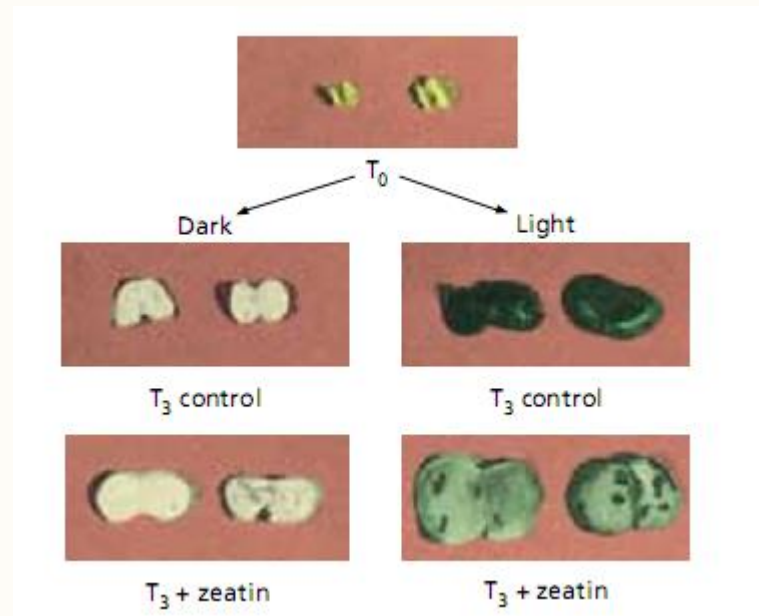
Leaf senescence is retarded in a transgenic tobacco plant containing a cytokinin biosynthesis gene, *ipt*.

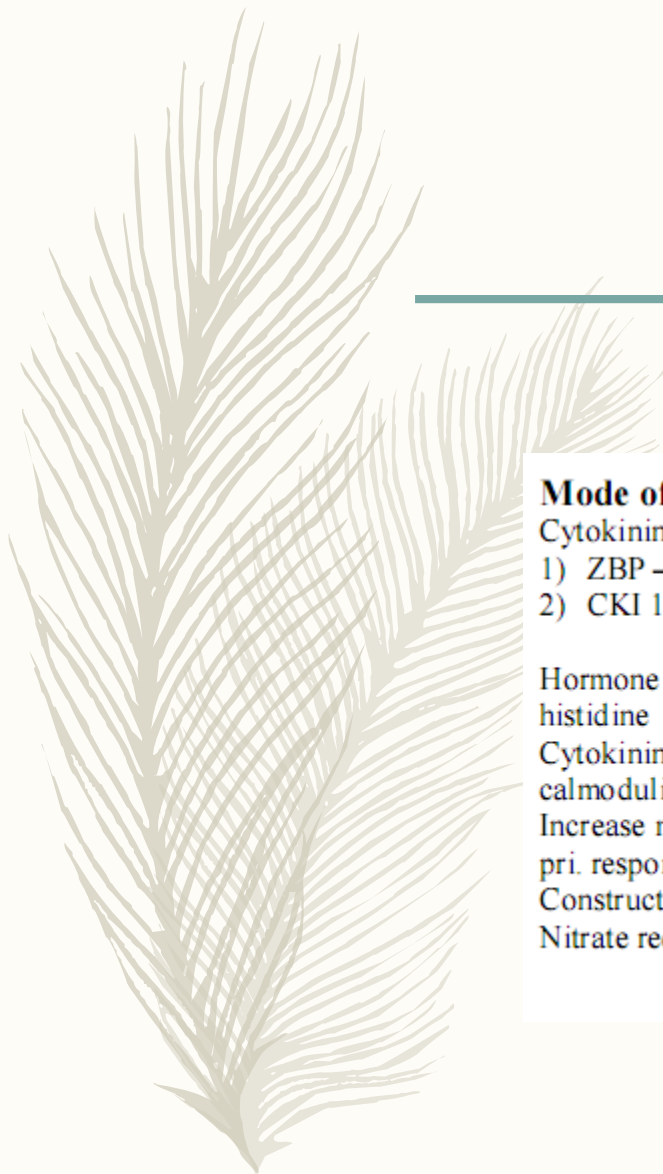


Plant expressing *ipt* gene remains green and photosynthetic

Age-matched control: advanced senescence, no photosynthesis

The effect of cytokinin on the expansion of radish cotyledons.





Mode of Action

Cytokinin receptor

- 1) ZBP – 67 Kda - high affinity for zeatin
- 2) CKI 1 (Cytokinin independent 1) – 125 Kda (similar to ETR 1)

Hormone binding → induce histidine kinase activity → autophosphorylation of histidine

Cytokinin signal transduction involves Ca^{2+} as a secondary messenger (Ca^{2+} and calmodulin regulated protein kinases)

Increase mRNA level- soyabean 20 different mRNA's increase 2-20 fold within 4 h (of pri. responsive genes)

Construction of cDNA libraries from cytokinin treated and starved tissues

Nitrate reductase induced by cytokinin (partially substitutes for light)

1. Cytokinin binds to CRE1, which is likely to occur as a dimer. Cytokinin binds to an extracellular portion of CRE1 called the CHASE domain. Two other hybrid sensor kinases (AHK2 and AHK3) containing a CHASE domain are also likely to act as cytokinin receptors in *Arabidopsis*.

2. Cytokinin binding to these receptors activates their histidine kinase activity. The phosphate is transferred to an aspartate residue (D) on the fused receiver domains.

3. The phosphate is then transferred to a conserved histidine present in an AHP protein.

4. Phosphorylation causes the AHP protein to move into the nucleus, where it transfers the phosphate to an aspartate residue located within the receiver domain of a type-B ARR.

5. The phosphorylation of the type-B ARR activates the output domain to induce transcription of genes encoding type-A ARRs.

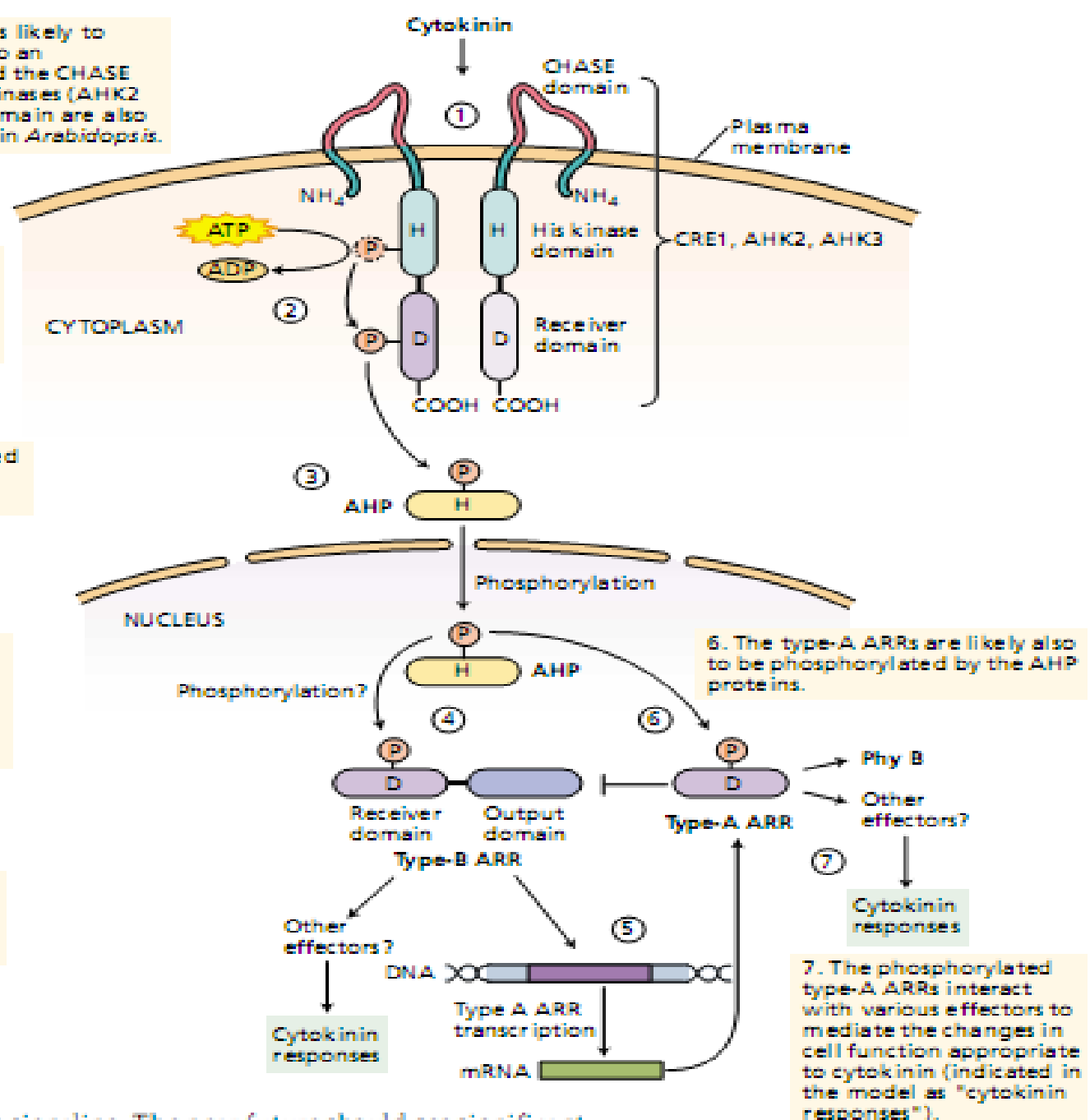


FIGURE 21.27 Model of cytokinin signaling. The near future should see significant refinement of this model, the tools are now in hand to analyze the interactions among these elements.

Gibberellins

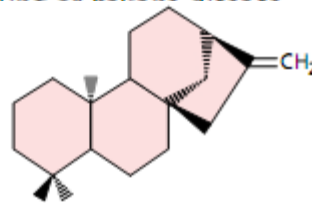
Discovery

Japan – first isolation

Giberella

1930 - first

All GA a

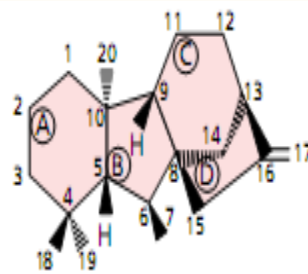


ent-Kaurene

are compounds –Giberellin A & B
structure

1958 – Jake Mac Millan – GA A₁ from a higher plant (seeds of *Phaseolus*) – seeds are better source

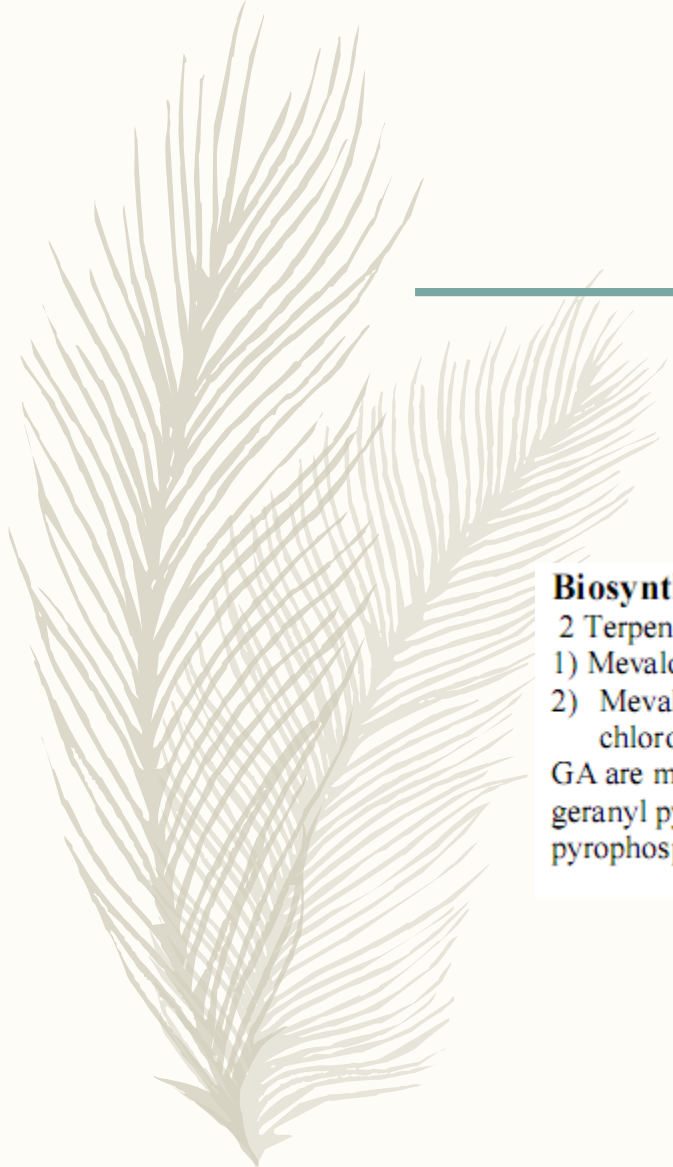
Numbered as GA_x in the order of their discovery (implies no close chemical similarity or metabolic reactions)



ent-Gibberellane structure

t-giberellane structure

e C-19



Biosynthesis

2 Terpenoid pathway

- 1) Mevalonic acid dependent pathway (precursor-mevalonic acid, in cytosol)
- 2) Mevalonic acid independent pathway (precursor-GLAL-3-P & pyruvate, in chloroplasts)

GA are made up of 4 isoprene units – isoprene units are added successively to produce geranyl pyrophosphate (C10), farnesyl pyrophosphate (C 15) and geranylgeranyl pyrophosphate (C 20)

GA synthesis has 3 stages

Stage 1: Cyclization reactions

GGPP \longrightarrow ent – kaurene

2 enzs. Are localized in the proplastids of meristematic shoots

AMO-1618, cycocel & Phosphon D are the specific inhibitors of this stage

Stage 2:

A methyl group oxidized to carboxylic group. Also one of the ring (B ring) contracts from a 6C to 5C ring – GA₁₂ aldehyde (1st GA formed and precursor to all other GA's)

Enzs. Localized in ER

Paclobutrazol inhibits this stage

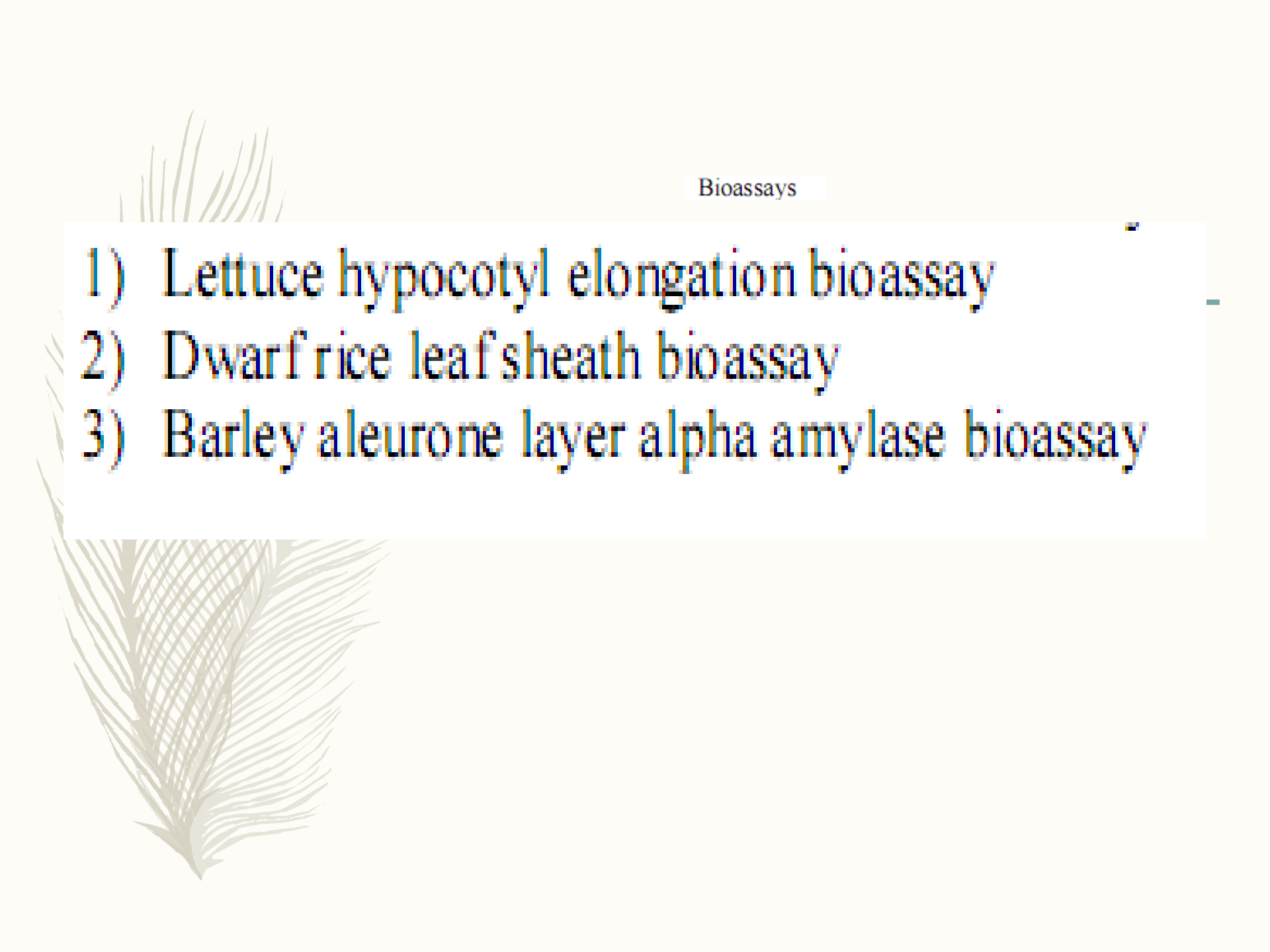
Stage 3:

All steps catalyzed by enz. dioxygenase located in the cytosol

2 basic chemical changes

1) hydroxylation at C-13 or C-3 or both

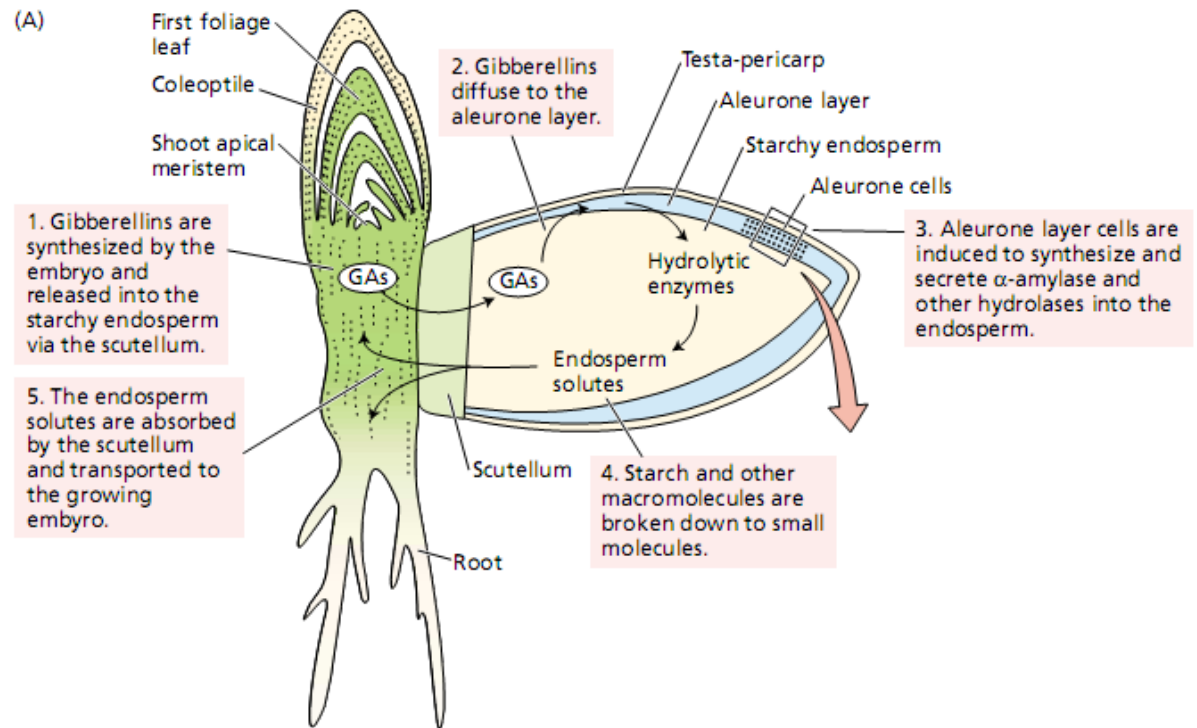
2) A successive oxidation at C-20 ($\text{CH}_2 \rightarrow \text{CH}_2\text{OH} \rightarrow \text{CHO}$), followed by a C loss of C-20




Bioassays

- 1) Lettuce hypocotyl elongation bioassay
- 2) Dwarf rice leaf sheath bioassay
- 3) Barley aleurone layer alpha amylase bioassay

(A)



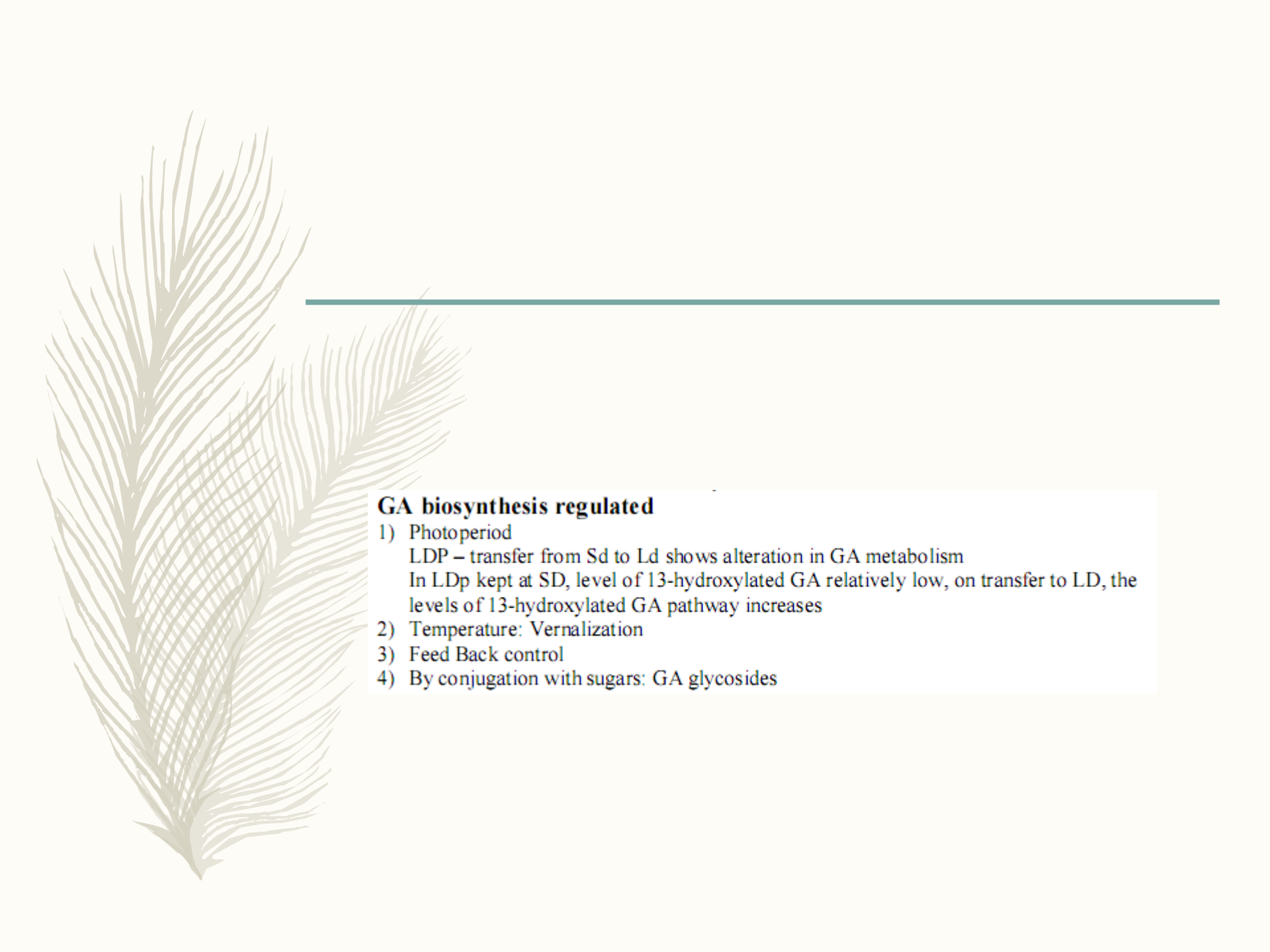


GA intermediates can be transported

High in developing seeds - decreases in mature seeds - mature seeds contain GA₁₂ aldehyde (the immediate GA precursor) - GA synthesis at young, growing buds, leaves & upper internodes – transported via phloem.


Initial steps of GA biosynthesis may occur in one tissue and metabolism to active GA in another

Ex. Intermediates of GA are transported from meristematic shoot tissues to green leaves



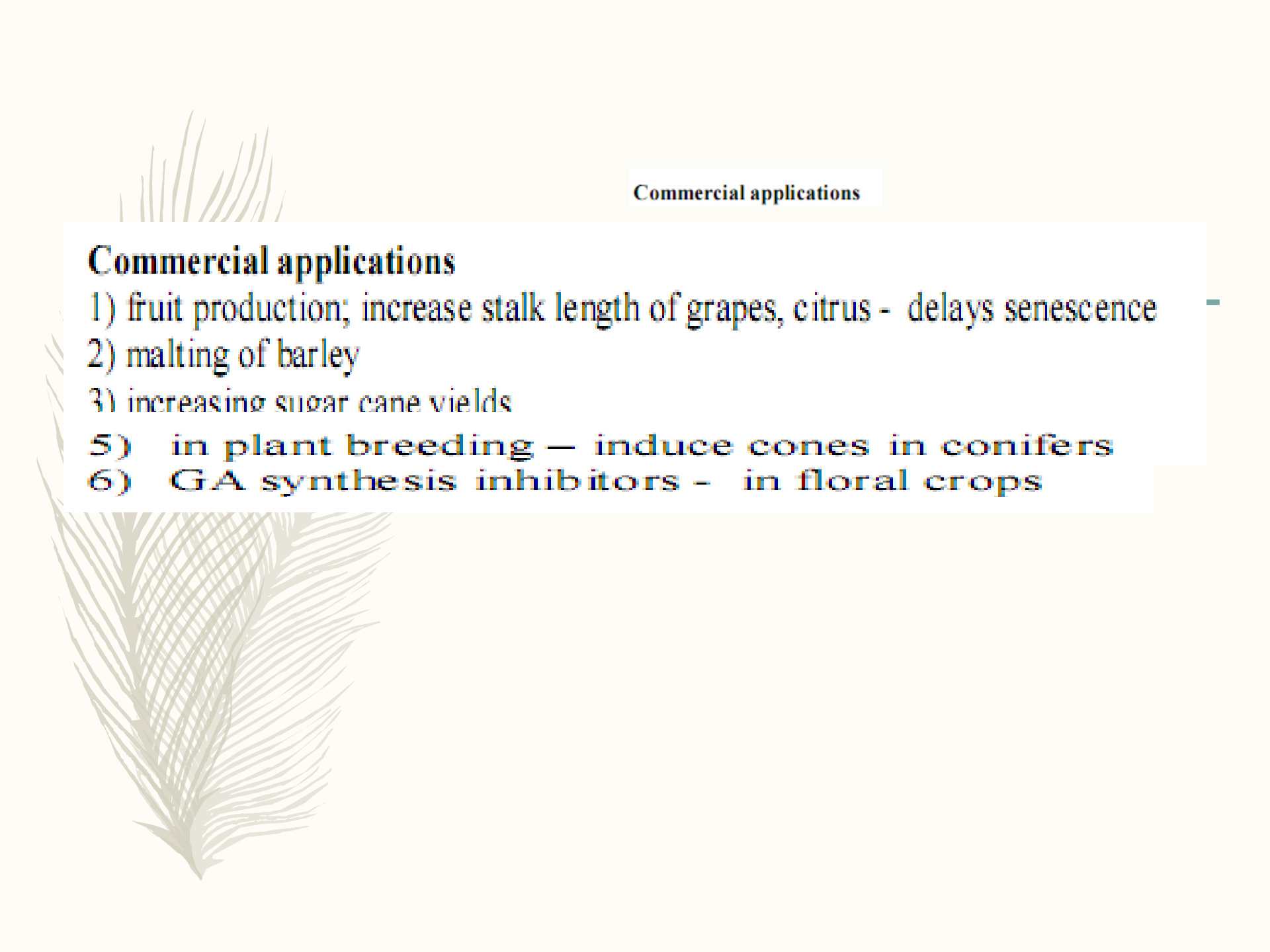
GA biosynthesis regulated

- 1) Photoperiod
LDP – transfer from Sd to Ld shows alteration in GA metabolism
In LDp kept at SD, level of 13-hydroxylated GA relatively low, on transfer to LD, the levels of 13-hydroxylated GA pathway increases
- 2) Temperature: Vernalization
- 3) Feed Back control
- 4) By conjugation with sugars: GA glycosides



Effects of GA on growth and development

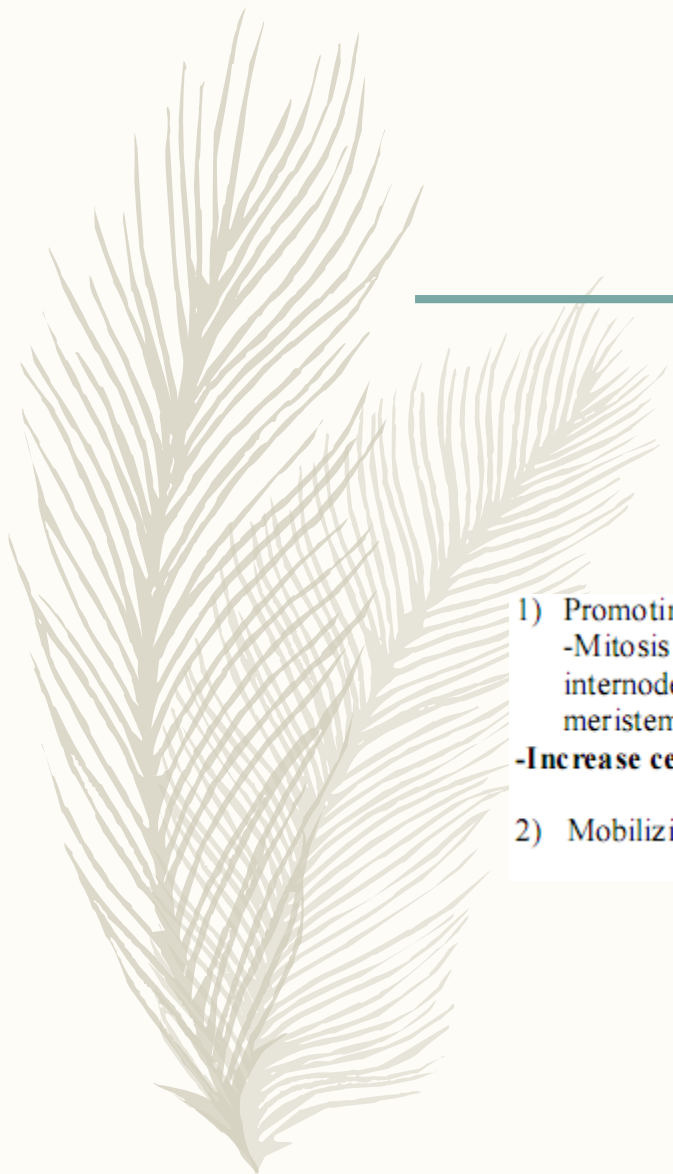
- 1) stimulates stem growth in dwarf and rosette plants (target of GA action in the intercalary meristem)
- 2) regulates transition from juvenile to adult phase
- 3) influence floral initiation and sex determination
- 4) promotes fruit set (ex. apple)
- 5) promotes seed germination



Commercial applications

Commercial applications

- 1) fruit production; increase stalk length of grapes, citrus - delays senescence -
- 2) malting of barley
- 3) increasing sugar cane yields
- 5) in plant breeding — induce cones in conifers
- 6) GA synthesis inhibitors - in floral crops



Mechanism of GA action

- 1) Promoting stem growth
 - Mitosis increases markedly in the subapical region of the meristem. Stimulation of internode elongation is partly due to increased cell division activity in the intercalary meristem. Regulates the cell cycle in intercalary meristem
 - Increase cell wall extensibility: XET - xyloglucan endotransglycosylase (activity increases), facilitates the penetration of expansins into the cell wall**
- 2) Mobilizing endosperm reserves

Mode of action

Receptors are localized on the membrane

GA – receptor – interaction with G protein - stimulates GDP/GTP exchange by heterotrimeric G proteins - induces the synthesis & secretion of alpha amylase gene.

1. GA₁ from the embryo first binds to a cell surface receptor.

2. The cell surface GA receptor complex interacts with a heterotrimeric G-protein, initiating two separate signal transduction chains.

3. A calcium-independent pathway, involving cGMP, results in the activation of a signalling intermediate.

4. The activated signalling intermediate binds to DELLA repressor proteins in the nucleus.

5. The DELLA repressors are degraded when bound to the GA signal.

6. The inactivation of the DELLA repressors allows the expression of the MYB gene, as well as other genes, to proceed through transcription, processing, and translation.

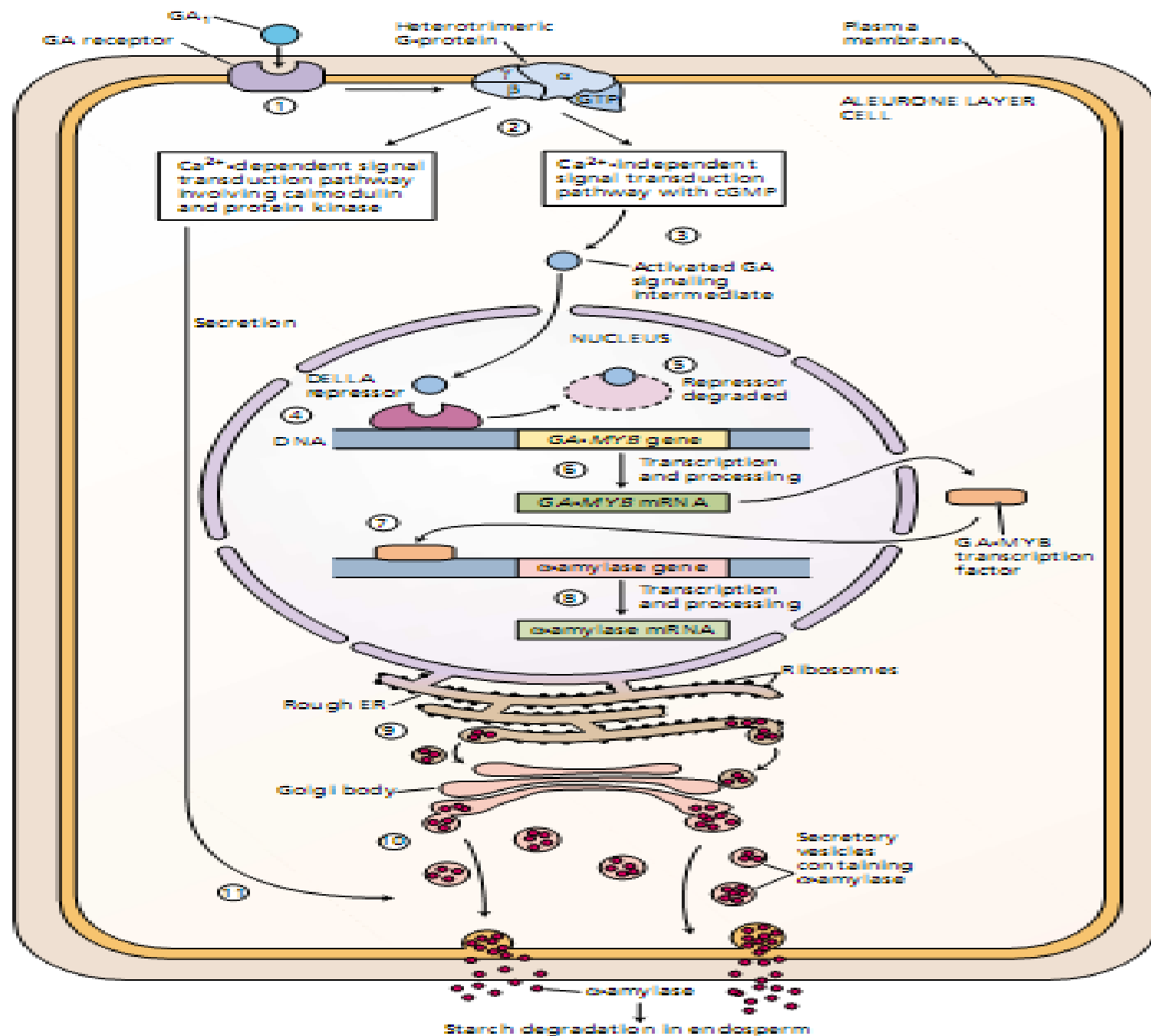
7. The newly synthesized MYB protein then enters the nucleus and binds to the promoter genes for α-amylase and other hydrolytic enzymes.

8. Transcription of α-amylase and other hydrolytic genes is activated.


9. α-Amylase and other hydrolases are synthesized on the rough ER.

10. Proteins are secreted via the Golgi.

11. The secretory pathway requires GA stimulation via a calcium-calmodulin-dependent signal transduction pathway.



Ethylene




19th century – coal gas – illumination, street lamps
Dimitry (Graduate student at St. Petersburg) – 1901 - triple response in pea seedlings (reduced stem elongation, increased lateral growth and abnormal horizontal growth)
1910 - Cousins – oranges – premature ripening of bananas

Biosynthetic regions and period

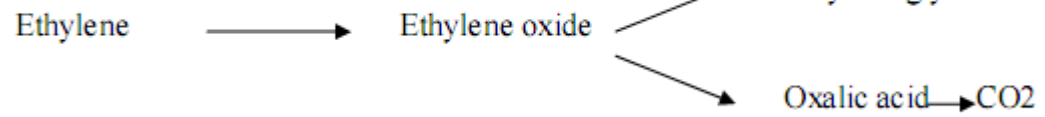
Meristematic and nodal regions are most active

- during fruit ripening, leaf senescence, abscission, wounding, stresses



Properties

Lighter than air, inflammable



Eth trapping system - KMnO_4

Biosynthesis

Methionine \longrightarrow Eth

AdoMet (S-adenosyl methionine) \longrightarrow ACC (1-aminocyclopropane-1-carboxylic acid)

ACC
Synthase

(regulated by envtal and internal factors, labile (unstable), purified using antisera, expressed in e. coli, showed similarity to aminotransferase superfamily of enzyme, encoded by multigene family)

ACC \longrightarrow Eth

ACC
oxidase

(members of Fe²⁺/ascorbate oxidase family)



Conjugated forms

N-malonyl ACC

GACC (1-(γ -L-glutamyl-amino) cyclopropane – 1- carboxylic acid

ACC synthase gene---antisense DNA

Eth production

- 1) Fruit ripening - \rightarrow ACC, ACC oxidase, ACC synthase
- 2) Stress induced – wounding, chilling, flooding, drought - \rightarrow Acc synthase
- 3) **Auxin induced - \rightarrow ACC synthase**

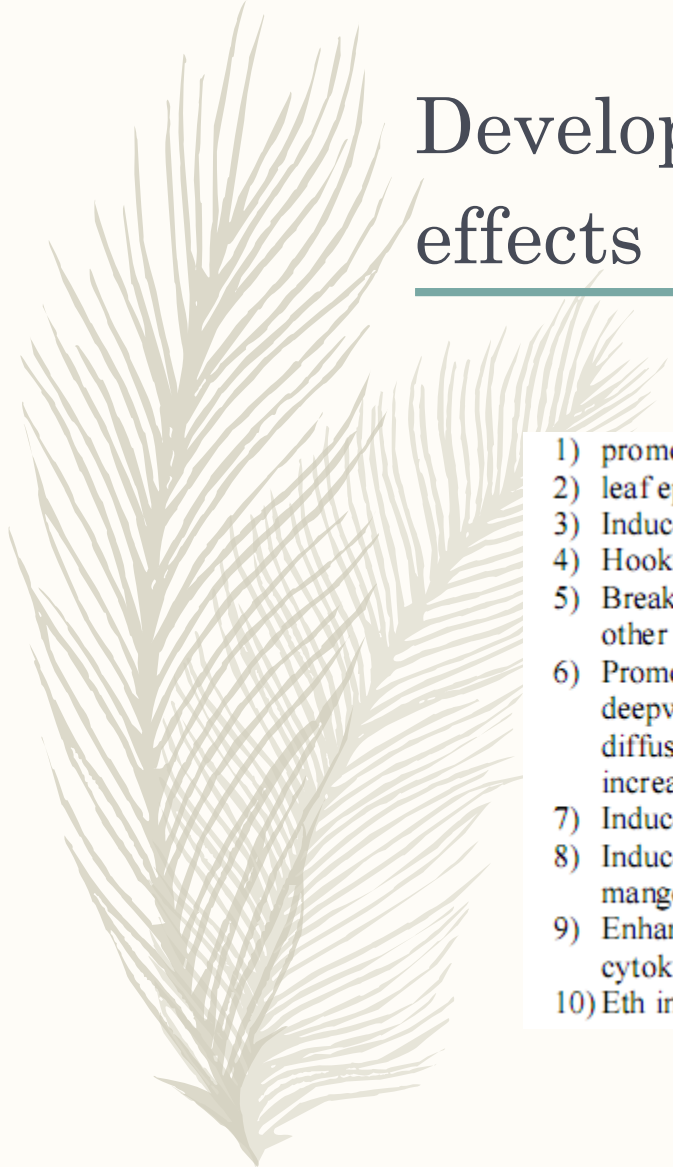
Inhibitors of Eth production

Adomet \longrightarrow ACC (inhibited by AVG (aminoethoxyvinylglycine), AOA
(aminooxyacetic acid)

ACC \longrightarrow Eth (inhibited by CO^{2+})

Eth action inhibited by Ag^+ of AgNO_3 , CO_2 (5-10%)

Trans-cyclooctene, competitive inhibitor to the ethylene for binding to the receptor



Developmental and physiological effects

- 1) promotes fruit ripening
- 2) leaf epinasty - when ACC from root is transported to shoot
- 3) Induces lateral cell expansion – triple response – pea
- 4) Hooks of dark grown seedlings - interaction between eth and phytochrome
- 5) Breaks seed and Bud dormancy - cereals, peanuts, sprouting in potato tubers and other bulbs
- 6) Promotes elongation of submerged aquatic sps. - ranunculus, nymphodes, deepwater rice (in absence of O₂ eth synthesis is diminished but loss of eth by diffusion is retarded. In deep water rice eth stimulates internode elongation by increasing sensitivity of cells of intercalary meristem to GA)
- 7) Induce root and root hairs (High concentration required - 10 µl/l)
- 8) Induce flowering in pineapple (synchronization of fruit set, flowering initiation in mango, promotion of female flowers in cucumber)
- 9) Enhances leaf senescence rate (senescence regulated by balance of eth and cytokinin)
- 10) Eth in abscission zone is regulated by Auxin

Triple response of etiolated pea seedlings. seedlings show a radial swelling, inhibition of elongation of the epicotyl, and horizontal growth of the epicotyl (diageotropism).



(A)



Epinasty, or downward bending of the tomato leaves. Epinasty results when the cells on the upper side of the petiole grow faster than those on the bottom.



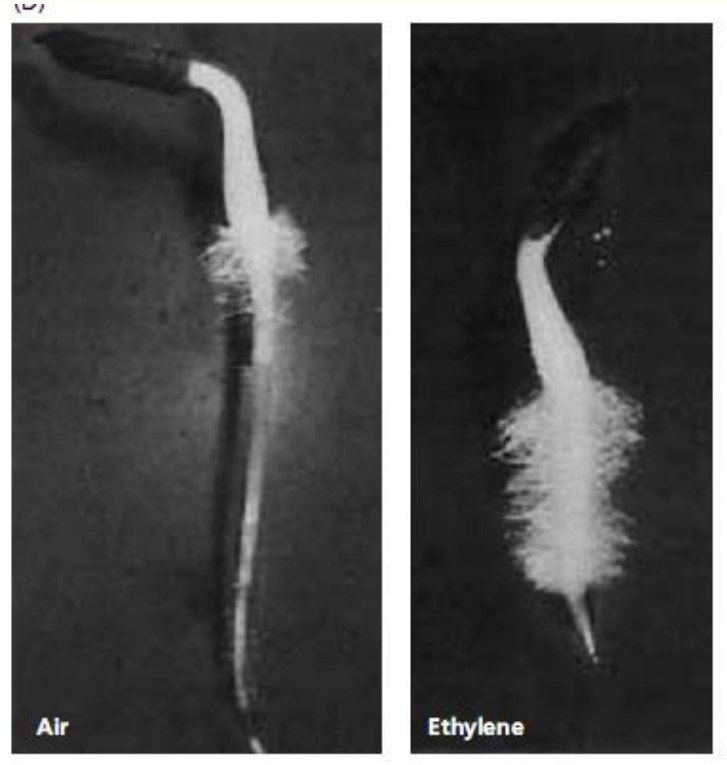
(B)



Inhibition of flower senescence by inhibition of ethylene action. Carnation flowers were held in deionized water for 14 days with (left) or without (right) silver thiosulfate (STS), a potent inhibitor of ethylene action. Blocking of ethylene results in a marked inhibition of floral senescence.



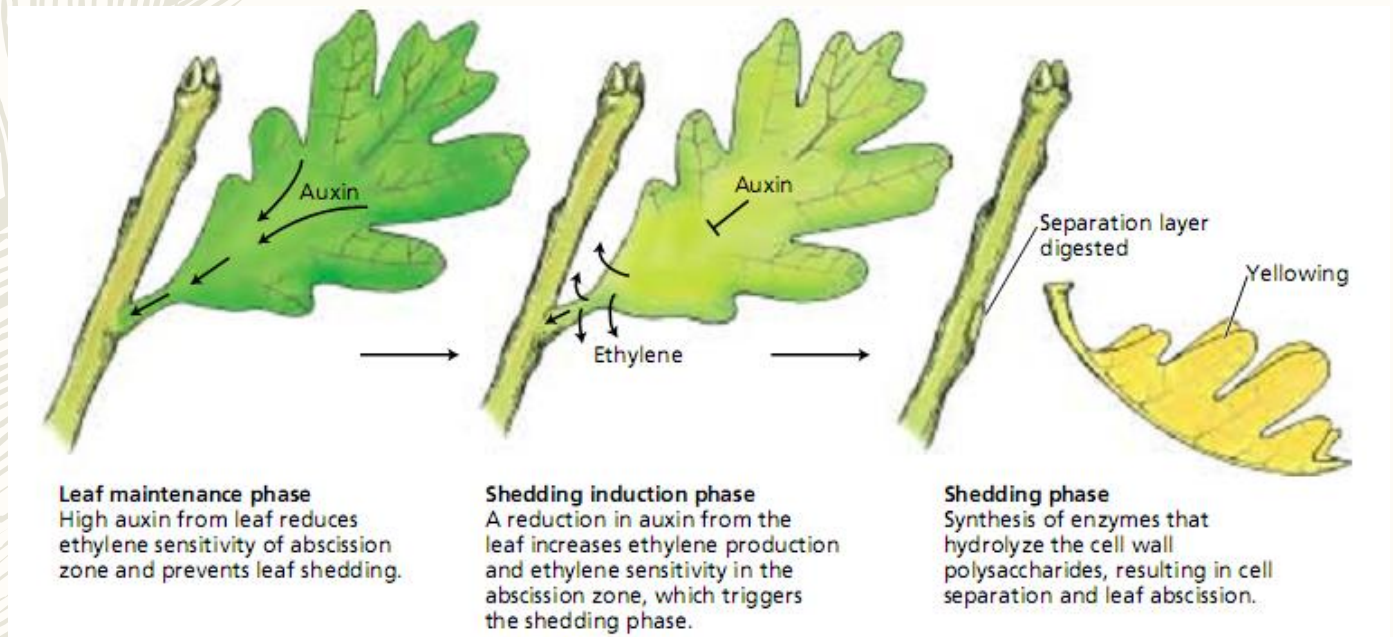
Promotion of root hair formation by ethylene in lettuce seedlings.



Effect of ethylene on abscission in birch (*Betula pendula*). The plant on the left is the wild type; the plant on the right was transformed with a mutated version of the *Arabidopsis* ethylene receptor, ETR1-1.



Schematic view of the roles of auxin and ethylene during leaf abscission.



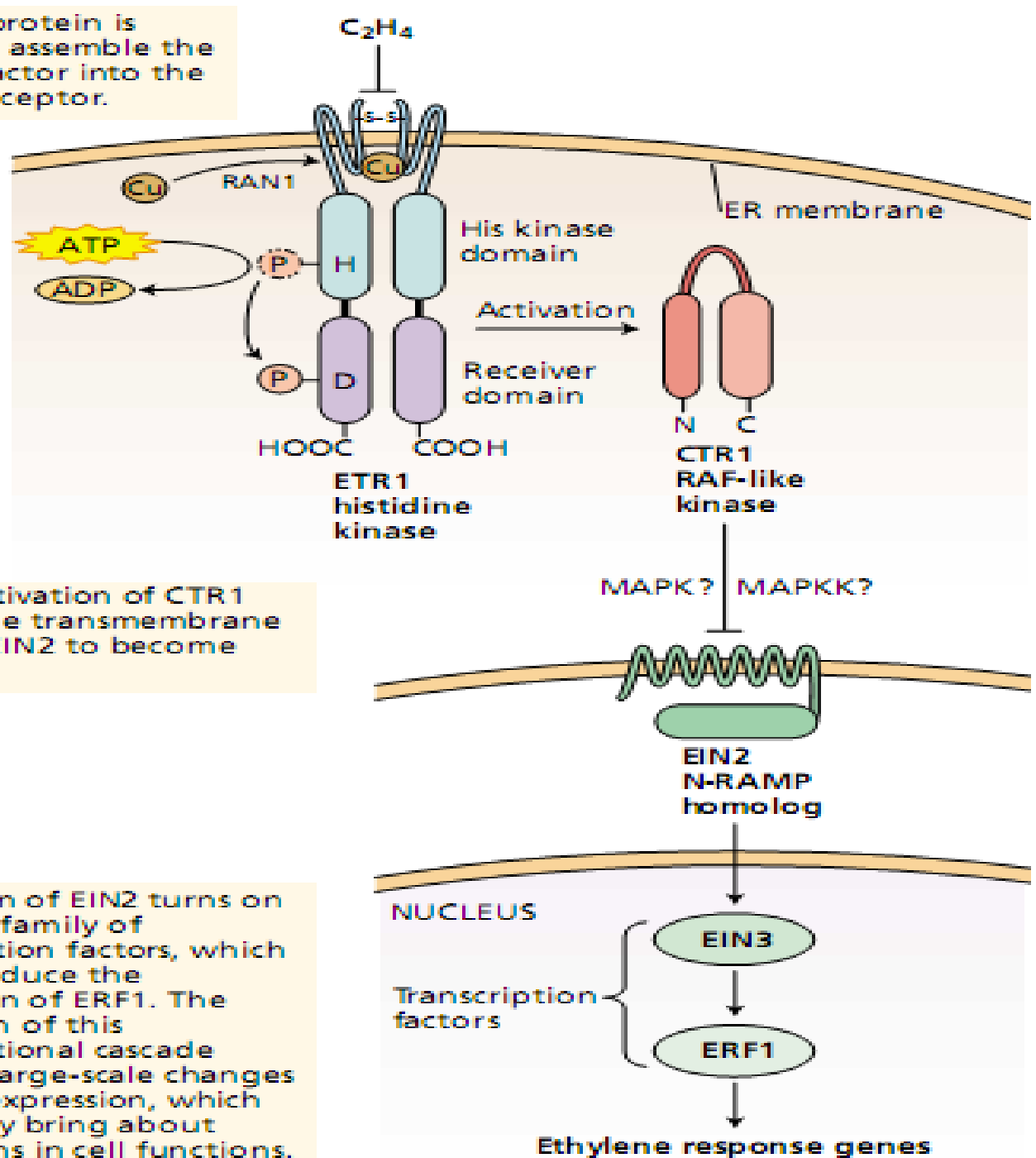
The RAN1 protein is required to assemble the copper cofactor into the ethylene receptor.

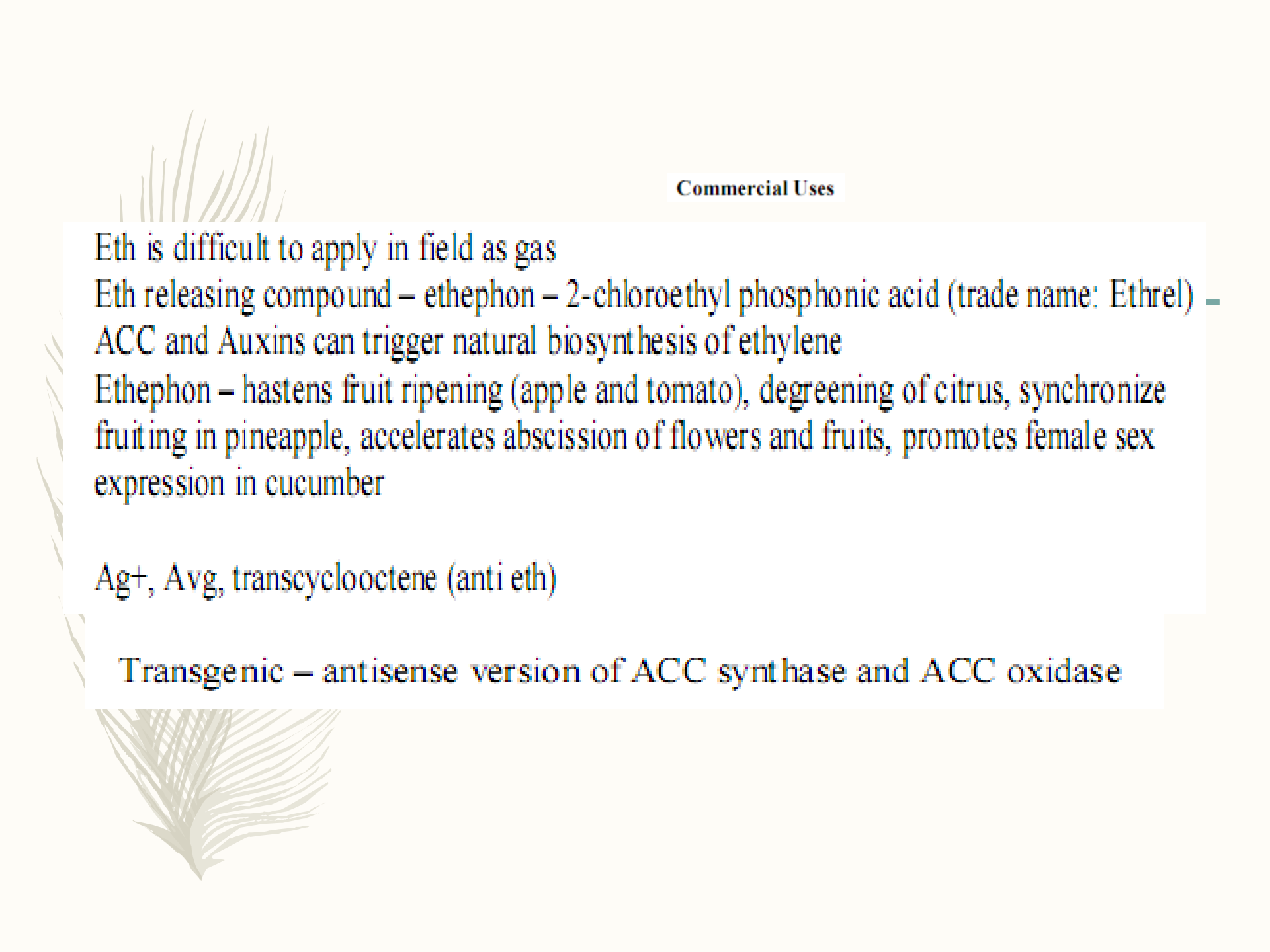
In the absence of ethylene, ETR1 and the other ethylene receptors activate the kinase activity of CTR1. This leads to a repression of the ethylene response pathway, possibly through a MAP kinase cascade. The binding of ethylene to the ETR1 dimer results in its inactivation, which causes CTR1 to become inactive.

The inactivation of CTR1 allows the transmembrane protein EIN2 to become active.

signaling in ETR1 receptor protein of plants of ethyl-cell; only receptor is a binds. membrane or, which is factors through

Activation of EIN2 turns on the EIN3 family of transcription factors, which in turn induce the expression of ERF1. The activation of this transcriptional cascade leads to large-scale changes in gene expression, which ultimately bring about alterations in cell functions.





Commercial Uses

Eth is difficult to apply in field as gas


Eth releasing compound – ethephon – 2-chloroethyl phosphonic acid (trade name: Ethrel) –

ACC and Auxins can trigger natural biosynthesis of ethylene

Ethephon – hastens fruit ripening (apple and tomato), degreening of citrus, synchronize fruiting in pineapple, accelerates abscission of flowers and fruits, promotes female sex expression in cucumber

Ag⁺, Avg, transcyclooctene (anti eth)

Transgenic – antisense version of ACC synthase and ACC oxidase



Mode of Action

Eth – receptor—signal transduction pathway – protein

Increases mRNA levels – for cellulase, chitinase, B-1,3-glucanase, peroxidase, chalcone synthase (key enz. in flavanoid synthesis), a pathogenesis related protein, ripening related genes, synthesis related genes

Eth responsive elements (ERE's)

- regulatory sequence – GCCGCC repeat motif

- ERE binding proteins (EREBP's) (eth pri. response gene products)

Fruit ripening – regulated by eth dependent (lycopene & aroma biosynthesis, respiratory metabolism and ACC synthase & independent pathways (chlorophyllase, ACC oxidase, polygalactouranase)